Quantitative Microbiology: Exponential Growth is Rarely Balanced

Rainer Machné

May 15, 2024

A Tale of Cell Biology, Told by Budding Yeast (and a Cyanobacterium)

A lecture series beyond the **known knowns** of (cell) biology, exploring the **known unknows**, the **unknown unknowns**, ... and some **unknown knowns** ...

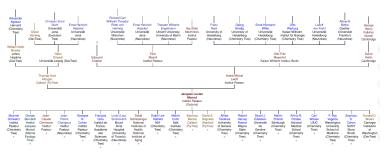
- 0. Quantitative Microbiology: Exponential growth is rarely balanced.
 - I. Pervasive transcription during the low energy phase of respiratory oscillations.
 - II. Transcription at LTR retrotransposons. TRANSCRIPTION
 - III. DNA as a metabolic sensor, and
 - IV. Chromosomal domains and mobile elements. GENOME HOMEOSTASIS
 - V. Protein homeostasis by a transcriptional oscillator, and
 - VI. Pulse-width modulation of gene expression. PROTEOME HOMEOSTASIS
 - VII. Metabolism: feedbacks and the auto-catalytic cycles of life, and
- VIII. The cell growth cycle as a cell-structural proofreading loop. METABOLISM
 - IX. Same, same in a cyanobacterium (circadian DNA supercoiling homeostasis).
 - X. Other eukaryotes: circadian and developmental clocks. OTHER SPECIES

Discussion: Do yeast cells dream of metabolic sheep?

A Tale of Cell Biology, Told by Budding Yeast (and a Cyanobacterium)

A lecture series beyond the **known knowns** of (cell) biology, exploring the **known unknows**, the **unknown unknowns**, ... and some **unknown knowns**

0. Quantitative Microbiology: Exponential growth is rarely balanced.



https://academictree.org (2023-09-26) of Jacques Monod

In this lecture series, we will trace a few specific paths through the vast directed acyclic graph of scientific progress. Is it a linear and logical process (Karl Popper) or a historical development, shaped by traditions, fashions and big egos (Ludwik Fleck)?

 $^{^{1} \}texttt{https://www.salatino.org/wp/are-citation-networks-really-acyclic/: "simple proof [\dots] we created a loop"}$

A Tale of Cell Biology, Told by Budding Yeast (and a Cyanobacterium)

A lecture series beyond the **known knowns** of (cell) biology, exploring the **known unknows**, the **unknown unknowns**, ... and some **unknown knowns** ...

0. Quantitative Microbiology: Exponential growth is rarely balanced.



©Yui Hachiya, https://artyui.happyforever.com/

In this lecture series, we will trace a few specific paths through the vast directed acyclic² graph of scientific progress. Is it a linear and logical process (Karl Popper) or a historical development, shaped by traditions, fashions and big egos (Ludwik Fleck)?

https://www.salatino.org/wp/are-citation-networks-really-acyclic/: "simple proof [...] we created a loop"















Saccharomyces cerevisiae: sugar-fungus of beer

- ightharpoonup Cultivated by and co-evolving with humans since \geq 5000 years.
- ▶ Plinius the Elder: $sugar + yeast \Rightarrow alcohol + more yeast$.
- Greek: ζνμη ~ dough, Latin: fermentum (hot, boiling),
 German/English: Hefe ~ raiser, Gischt/Gest ~ yeast,
 Austrian: Germ ~ gären or germen (Latin) ~ germ?



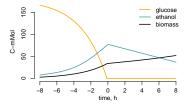












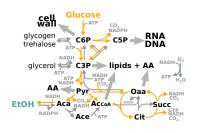
'The alcohol level is [...] about 14%. Aroma has a great spicy orange edge to it. Review coming.' http://beerobsessed.com/blog/?p=1044 (2017)

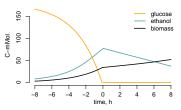
Zampar et al. (2013) Mol Syst Biol

Saccharomyces cerevisiae: sugar-fungus of beer

- ▶ Leeuwenhoek: contains cells, Schwann (1837): a single celled fungus!
- Louis Pasteur vs. Justus von Liebig: Is alcoholic fermentation performed by a life force or by a chemical catalyst?
- Development of minimal and defined growth media (minerals+sugar), and discovery of vitamins (Lindner (1919): the search for bios).

$$\xrightarrow{\text{CH}_2\text{OH}} + \xrightarrow{\text{OH}} + \bigoplus_{\text{OH}} + \bigoplus_{\text{CO}_2} + \bigoplus_{\text{H}_1} + \bigoplus_{\text{H}_2} + \bigoplus_{\text{CO}_2} + \bigoplus_{\text{C}_2} +$$





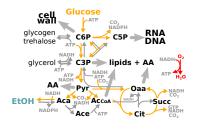
Zampar et al. (2013) Mol Syst Biol mammals: various from Rabinowitz Lab

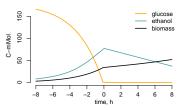
en-zyme: 'in yeast'

also in cancer (lactate)!

- ▶ Both Pasteur and Liebig were right: Microbiology and Biochemistry.
- Pasteur effect: more alcohol without oxygen, Warburg/Crabtree effects: fermentation even in the presence of oxygen.
- At high sugar concentration, yeast first ferments sugar to alcohol, then consumes alcohol for a second growth phase (diauxic).

$$\xrightarrow{\text{CH}_2\text{OH}} + \qquad \xrightarrow{\text{ethanol}_1^H} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{sugar}_{\text{OH}}} + \qquad \xrightarrow{\text{h}_{\text{-}}^H} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{sugar}_{\text{OH}}} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{sugar}_{\text{OH}}} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{$$





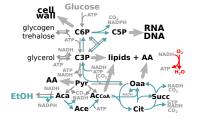
Zampar et al. (2013) Mol Syst Biol mammals: various from Rabinowitz Lab

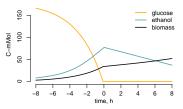
en-zyme: 'in yeast'

also in cancer (lactate)!

- ▶ Both Pasteur and Liebig were right: Microbiology and Biochemistry.
- Pasteur effect: more alcohol without oxygen, Warburg/Crabtree effects: fermentation even in the presence of oxygen.
- At high sugar concentration, yeast first ferments sugar to alcohol, then consumes alcohol for a second growth phase (diauxic).

$$\xrightarrow{\text{CH}_2\text{OH}} + \qquad \xrightarrow{\text{ethanol}_1^H} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{sugar}_{\text{OH}}} + \qquad \xrightarrow{\text{ch}_2^H} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{sugar}_{\text{OH}}} + \qquad \xrightarrow{\text{co}_2} + \qquad \xrightarrow{\text{co}_2^H} + \qquad \xrightarrow{\text{c$$





Zampar et al. (2013) Mol Syst Biol mammals: various from Rabinowitz Lab

en-zyme: 'in yeast'

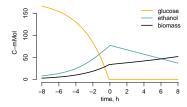
also in cancer (lactate)!

- ▶ Both Pasteur and Liebig were right: Microbiology and Biochemistry.
- Pasteur effect: more alcohol without oxygen, Warburg/Crabtree effects: fermentation even in the presence of oxygen.
- At high sugar concentration, yeast first ferments sugar to alcohol, then consumes alcohol for a second growth phase (diauxic).

$$\xrightarrow{\text{CH}_2\text{OH}}_{\text{OH}} + \xrightarrow{\text{CO}_2}_{\text{OH}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{C}} + \xrightarrow{\text{CO}_2}_{\text{C}}$$

$$\begin{split} \frac{dX}{dt} &= \mu X \\ \frac{dS}{dt} &= S_0 - \frac{1}{y} \mu X \; , \end{split}$$

with growth rate $\mu = f(S)$ and yield $y = \frac{\Delta X}{-\Delta S}$.



Zampar et al. (2013) Mol Syst Biol Andrea Weisse et al. (2015) PNAS

Cells as auto-catalytic enzymes.

- Jacques Monod at the Institut Pasteur: from Physiology to Genetics.
- Diauxic Growth: E. coli and B. subtilis first use glucose, then other sugars, with lower yield y and/or lower growth rate μ.
- Discovery of gene regulation: the *lac* operon (with Francois Jacob)!

$$\xrightarrow{\text{ch}_2\text{OH}} + \xrightarrow{\text{oh}} + \bigoplus_{\text{Sugar}_{\text{OH}}} + \bigoplus_{\text{H}} + \bigoplus$$

$$egin{aligned} rac{dX}{dt} &= \mu X \ rac{dS}{dt} &= S_0 - rac{1}{y} \mu X \ \mu &= \mu_{ ext{max}} rac{S}{S + K_S}. \end{aligned}$$

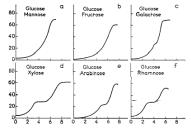


Fig.1. Growth of Esherichia coli in the presence of different carbohydrate pairs serving as the only source of carbon in a synthetic medium.

Monod (1942) Doctoral Thesis

Cells as auto-catalytic enzymes.

m Physiology to Constice

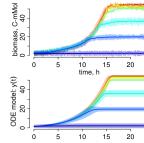
Quantitative Microbiology

- ▶ Jacques Monod at the Institut Pasteur: from Physiology to Genetics.
- Diauxic Growth: E. coli and B. subtilis first use glucose, then other sugars, with lower yield y and/or lower growth rate μ.
- Discovery of gene regulation: the *lac* operon (with Francois Jacob)!

Two steps forward, one back (y-axis labels!): absorbance ∞ biomass ∞ cell number?

$$\xrightarrow{\text{CH}_2\text{OH}} + \xrightarrow{\text{OH}} + \bigoplus_{\text{OH}} + \bigoplus_{\text{CH}_2\text{OH}} + \bigoplus_{\text{H}_2\text{OH}} + \bigoplus_{\text$$

$$egin{aligned} rac{dX}{dt} &= \mu X \ rac{dS}{dt} &= S_0 - rac{1}{y} \mu X \ \mu &= \mu_{ ext{max}} rac{S}{S + K_S}. \end{aligned}$$



E. coli at different glucose concentrations; BIQ980, 2019.

Cells as auto-catalytic enzymes.

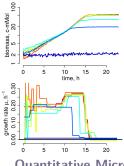
Quantitative Microbiology

- Jacques Monod at the Institut Pasteur: from Physiology to Genetics.
- Diauxic Growth: E. coli and B. subtilis first use glucose, then other sugars, with lower yield y and/or lower growth rate μ.
- Discovery of gene regulation: the *lac* operon (with Francois Jacob)!

Two steps forward, one back (y-axis labels!): absorbance ∞ biomass ∞ cell number?

$$\xrightarrow{\text{CH}_2\text{OH}} + \xrightarrow{\text{OH}} + \bigoplus_{\text{OH}_2\text{OH}} + \bigoplus_{\text{CH}_2\text{OH}} + \bigoplus_{\text{H}_2\text{H}_2\text{OH}} + \bigoplus_{\text{H}_2\text{H}_2\text{OH}_$$

$$egin{aligned} rac{dX}{dt} &= \mu X \ rac{dS}{dt} &= S_0 - rac{1}{y} \mu X \ \mu &= \mu_{ ext{max}} rac{S}{S + K_S}. \end{aligned}$$



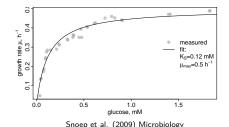
Cells as auto-catalytic enzymes.

- **Quantitative Microbiology**
- Jacques Monod at the Institut Pasteur: from Physiology to Genetics.
- Diauxic Growth: E. coli and B. subtilis first use glucose, then other sugars, with lower yield y and/or lower growth rate μ.
- Discovery of gene regulation: the *lac* operon (with Francois Jacob)!

Nice algorithm: piecewise linear segmentation by dpseg, Machné and Stadler (2019) CRAN.

$$\xrightarrow{\text{CH}_2\text{OH}} + \xrightarrow{\text{OH}} + \bigoplus_{\text{OH}} + \bigoplus_{\text{CH}_2\text{OH}} + \bigoplus_{\text{H}} + \bigoplus_{\text{H}} + \bigoplus_{\text{H}} + \bigoplus_{\text{H}} + \bigoplus_{\text{C}} + \bigoplus_{\text$$

$$egin{aligned} rac{dX}{dt} &= \mu X - \phi X \ rac{dS}{dt} &= S_0 - rac{1}{y} \mu X - \phi S \ \mu &= \mu_{ ext{max}} rac{S}{S + K_S}. \end{aligned}$$

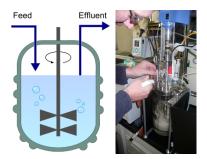


Cells as auto-catalytic enzymes.

- Jacques Monod at the Institut Pasteur: from Physiology to Genetics.
- ▶ Diauxic Growth: *E. coli* and *B. subtilis* first use glucose, then other sugars, with lower yield y and/or lower growth rate μ .
- Discovery of gene regulation: the *lac* operon (with Francois Jacob)!
- La culture continue (Monod 1950) or the chemostat (Novick and Szilard 1950).

$$\xrightarrow{\text{ch},\text{OH}}_{\text{OH}} + \xrightarrow{\text{ch}}_{\text{OH}} + \xrightarrow{\text{ch}}_{\text{OH}} + \xrightarrow{\text{ch},\text{OH}}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{C}} + \xrightarrow{\text{ch},\text{OH}}_{\text{C}}$$

$$rac{dX}{dt} = \mu X - \phi X$$
 $rac{dS}{dt} = S_0 - rac{1}{y}\mu X - \phi S$ $\mu = \mu_{\mathsf{max}} rac{S}{S + K_S}.$



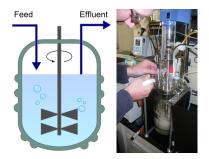
Cells as auto-catalytic enzymes.

- Jacques Monod at the Institut Pasteur: from Physiology to Genetics.
- ▶ Diauxic Growth: *E. coli* and *B. subtilis* first use glucose, then other sugars, with lower yield y and/or lower growth rate μ .
- Discovery of gene regulation: the *lac* operon (with Francois Jacob)!
- La culture continue (Monod 1950) or the chemostat (Novick and Szilard 1950).

$$\xrightarrow{\text{CH}_2\text{OH}}_{\text{OH}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{C}} + \xrightarrow{\text{Sugar}_{\text{OH}}}$$

At $\frac{dX}{dt} = 0$, the dilution rate sets the growth rate:

$$\phi = \frac{\mathrm{flow\ rate}}{\mathrm{volume}} = \mu = \frac{\ln 2}{\tau_{\mathrm{doubling}}}$$



Continuous Culture:

- ▶ Stable states of growth can be achieved over a large range of $\phi = \mu$.
- ▶ Cell growth is then limited by a single nutrient in the growth medium.

$$\xrightarrow{\text{CH}_2\text{OH}}_{\text{OH}} + \xrightarrow{\text{OH}}_{\text{OH}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{H}} + \xrightarrow{\text{CO}_2}_{\text{C}} +$$

Dilution rate is 1/retention time:

$$\phi =$$
 0.303 h $^{-1}$ $au_{
m doubling} = rac{\ln 2}{\phi} = 2.3$ h $pprox au_{
m osc}$

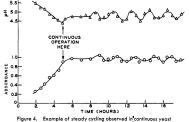


Figure 4. Example of steady cycling observed in continuous yeas propagator

Continuous Culture:

- Finn and Wilson (1954), Finn (1954): Saccharomyces carlsbergensis → Saccharomyces pastorianus³, at 2% glucose.
- ▶ Spontaneous synchronization of cell division, with a period τ_{osc} close to the doubling time $\tau_{doubling}$.
 - ⇒ (i) cells "communicate" and (ii) metabolic differences of cell cycle phases.

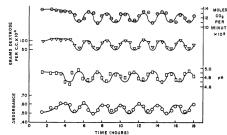
 $^{^3}$ old (~1k years) **bottom-fermenting hybrid** of *S. cerevisiae* and *S. eubayanus* (Libkind et al. 2011)

$$\xrightarrow{\text{CH}_2\text{OH}} + \bigoplus_{\text{OH}} + \bigoplus_{\text{CH}_2\text{$$

Figure 6. Growth of S. carlsbergensis in batch vessel at constant pH levels

Dilution rate is 1/retention time:

$$\phi=$$
0.303 h $^{-1}$ $au_{
m doubling}=rac{\ln 2}{\phi}=$ 2.3 h $pprox au_{
m osc}$



Continuous Culture:

Quantitative Microbiology

- Finn and Wilson (1954), Finn (1954): Saccharomyces carlsbergensis → Saccharomyces pastorianus, at 2% glucose.
- ▶ Spontaneous synchronization of cell division, with a period $\tau_{\rm osc}$ close to the doubling time $\tau_{\rm doubling}$, and
- ▶ with dynamic changes to culture pH and CO₂ production.

Exponential growth is rarely balanced ...

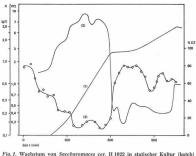
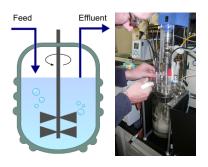


Fig. L. Wachstum von Soccharomgess eer. H 1922 in statischer Kultur (batch) in Medium N.10 (0.92% Glucos) unter aeroben Bedingungen. Anderungen der Trockensubstanz (X), des Respirationsquotienten (RQ) und der Populationsverteilung in Funktion der Zeit. Kurve (I): Trockensubstanz X. g./h, logarifhmisch aufgetragen. Kurve (2): Respirationsquotient RQ — $cog_0Q_{0.5}$ Kurve (B): $cog_0Q_{0.5}$ Kurve (B):



Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

Reproducible Biology: bioreactors with dissolved O₂ and pH measurement, and constant pH and temperature by PID controllers.

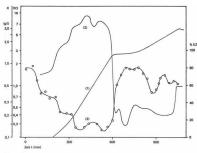
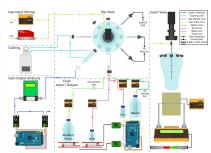


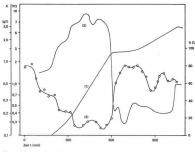
Fig.1. Wachstum von Saccharomyces cer. H
 1022 in statischer Kultur (batch) in Medium Ni. 10 (0.92% Glucos) unter aeroben Bedingungen. Amerungen der Trockensubstanz (X), des Respirationsquotienten (RQ) und der Populationstretikung in Prokukion der Zeli, Kuwre (I): Trockensubstanz X, gl., fügarühren verteilung in Funktion der Zeli, Kuwre (II): Trockensubstanz X, gl., fügarühren verteilung in Funktion der Zeli, Kuwre (II): Trockensubstanz X, gl., fügarühren verteilung in Funktion der Verteilung v



Behle, Dietsch et al. (2022) Nucl Acids Res Arduino-based offgas module (500 €), and weigh module (100 €).

Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

- Reproducible Biology: bioreactors with dissolved O₂ and pH measurement, and constant pH and temperature by PID controllers.
- Quantitative Biology: with controlled aeration (ingas) and offgas measurements (CO₂, O₂, H₂S) we have a quantitative picture of growth. We know what goes in (defined medium and gas) and what comes out.
- Including metabolic heat production: Calorimetry via $T_{
 m culture}-T_{
 m bath}$.



glycogen trehalose ATP NADPH SUCC.

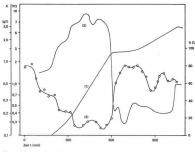
NADPH ACCO. NADPH ACCO. NADPH ACCO. NADPH ACCO. NADPH ACCO. STATE ACCO. NADPH ACCO. STATE ACCO. NADPH ACCO. STATE ACCO. STATE

Glucose

 $Fig.\,I.$ Wachstum von Saccharomyces eer. Il 1022 in statischer Kultur (hatch) in Medium N.I. of 0.925 Glucose) unter aeroben Bedingungen. Anderungen der Trockensubstanz (X), des Respirationsquotienten (RQ) und der Populationsverteilung in Funktion der Zeit. Kurve (1): Trockensubstanz X, gf., logarithmisch aufgetragen. Kurve (2): Respirationsquotient RQ — $Q_{\rm CQ}$, $Q_{\rm D}$, Kurve (3): $Q_{\rm CQ}$, where $Q_{\rm CQ}$ is the first of the first

Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

- ▶ Von Meyenburg (1968): dynamic RQ at constant biomass growth rate!
- ► The Respiratory Quotient: $RQ = \frac{q_{CO_2}}{-q_{O_2}}$:
 - 1. RQ \approx 1: complete oxidation of glucose to CO₂,
 - 2. RQ > 1: additional CO_2 by fermentation,
 - 3. RQ $\approx \frac{2}{3}$: complete oxidation of ethanol.
- EZ %: percent of non-budding cells.



 $Fig.\,1.$ Wachstum von Saccharomyees eer. Il 1022 in statischer Kullur (hatch) in Medium N.1. 0 (0.925 Glucose) unter aeroben Bedingungen. Anderungen der Trockensubstanz (X), des Respirationsquotienten (RQ) und der Populationsverteilung in Funktion der Zeit. Kurve (1): Trockensubstanz X, gf., logarithmisch aufgetragen. Kurve (2): Respirationsquotient RQ — $Q_{\rm CQ_2}$ /Qo., Kurve (3): 100% — % D.2. der der Einzelteilen (nicht sprossend) in der Population. % Zeit. 2 (1): 100% — % D.2.

Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

- ▶ Von Meyenburg (1968): dynamic RQ at constant biomass growth rate!
- ► The Respiratory Quotient: $RQ = \frac{q_{CO_2}}{-q_{O_2}}$:
 - 1. RQ \approx 1: complete oxidation of glucose to CO₂,
 - 2. RQ > 1: additional CO_2 by fermentation,
 - 3. RQ $\approx \frac{2}{3}$: complete oxidation of ethanol.
- ► EZ %: percent of non-budding cells.

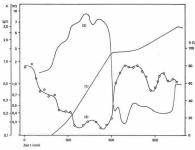
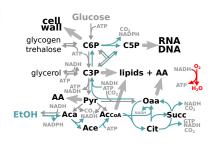


Fig. 1. Waehstum von Saccharomyces cer. H 1022 in statischer Kultur (bateh) in Medium Nl. 10 (9.92% Glucose) unter aeroben Bedingungen. Änderungen der Trockensubstanz (X), des Respirationsyudoitente (RQ) und der Populationsverteilung in Funktion der Zeit. Kurve (I): Trockensubstanz X, g/l, logarithemisch aufgetragen, Kurve (2): Sepsirationsyudoitent RQ — $C_{\rm CQ}({\rm QQ}, {\rm Kurve}~{\rm G})$: Prozentualer Anteil der Einzelzellen (nicht sprossend) in der Population. % EZ = 100% - % D/C.

Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich



- ▶ Von Meyenburg (1968): dynamic RQ at constant biomass growth rate!
- ► The Respiratory Quotient: $RQ = \frac{q_{CO_2}}{-q_{O_2}}$:
 - 1. RQ \approx 1: complete oxidation of glucose to CO₂,
 - 2. RQ > 1: additional CO_2 by fermentation,
 - 3. RQ $\approx \frac{2}{3}$: complete oxidation of ethanol.
- ► EZ %: percent of non-budding cells: change of phase durations!

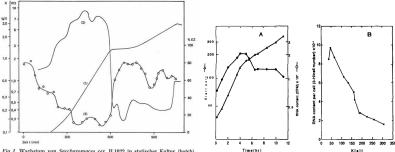
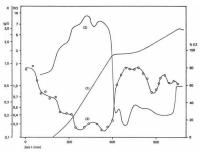


Fig. 1. Wachstum von Saccharomyces cer. H 1022 in statischer Kultur (batch) in Medium N.1 to 0,92% Glucosy unter aeroben Bedingungen. Anderungen der Trockensubstanz (N.) des Respirationsquotienten (RQ) und der Population verteilung in Funktion der Zeit. Kurre (II) Trockensubstanz N. giv. lioparitheren verteilung in Funktion der Zeit. Kurre (II) Trockensubstanz N. giv. lioparitheren verteilung in Funktion der Zeit. Kurre (II) Trockensubstanz N. giv. lioparitheren verteilung in Funktion der Population. Setz = 100% — 8 DZ.

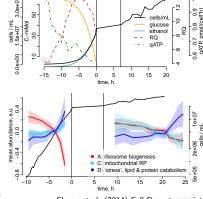
Figure 1. Ribosomal RNA content of growing yeast cell. A culture of W303a Ju and Warner (1994) Yeast

Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

- ▶ Von Meyenburg (1968): dynamic RQ at constant biomass growth rate!
- ▶ Ju and Warner (1994): ribosomal RNA per cell continuously decreases, cells sense the potential for growth?



 $Fig.\,I.$ Wachstum von Saccharomyees eer. II 1022 in statischer Kultur (batch) in Medium NI. 10 (0.925 Glucose) unter aeroben Bedingungen. Anderungen der Trockensubstanz (X), des Respirationsquotienten (RQ) und der Populationsverteilung in Funktion der Zeit. Kurve (1): Trockensubstanz & g.l. (agarithmisch aufgetragen. Kurve (2): Respirationsquotient RQ $= \mathrm{Qc_{20}}(\mathrm{Qo}_{2},\mathrm{Kurve}$ (3): $\mathrm{1005} - \mathrm{S}~\mathrm{D}.\mathrm{Ce}$ included der Einzettellen (nicht spressens) in der Population. SEZ



Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

Slavov et al. (2014) Cell Rep - transcripts

- ▶ Von Meyenburg (1968): dynamic RQ at constant biomass growth rate!
- ▶ Ju and Warner (1994): ribosomal RNA per cell continuously decreases.
- ▶ Slavov et al. (2014): dynamic RQ as above, qATP decreases
 - + omics: large transcript groups continuously change accordingly.

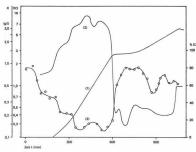
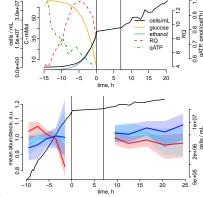


Fig. 1. Wachstum von Saccharomyces cer. H 1022 in statischer Kultur (batch) in Medium NL 10 (0.92% Glucose) unter aeroben Bedingungen. Anderungen der Trockensubstanz (X), des Respirationsquotienten (RQ) und der Populationsverteilung in Funktion der Zeit. Kurve (1): Trockensubstanz X, gl. logarithmisch aufgetragen. Kurve (2): Respirationsquotient RQ = $0_{\rm CQ}/0_{\rm Q}$, Kurve (3): Prozentualer Anteil der Einzelzellen (nicht sprossend) in der Population. % EZ = 100% - % DZ.



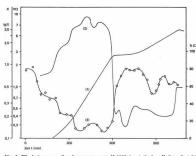
Slavov et al. (2014) Cell Rep - proteins

Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

.. which didn't matter for Molecular Biology and Genetics:

- An intersecting path through the DAG of scientific progress, incl. Lamarck/Darwin/Mendel, Morgan (PostDoc mentor of Monod), Schroedinger, Luria/Bertani (CSHL), Chargaff/Franklin, Watson/Crick, McClintock, Doudna/Charpentier, . . .
- Ignore the complex growth on LB medium, and just "harvest at OD=0.3".

Lectures II-IV



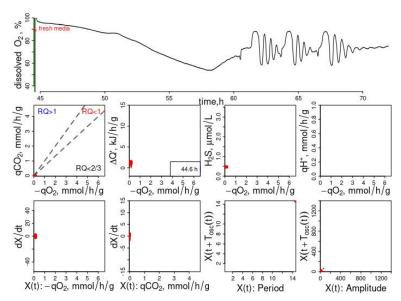
10 15 time. h mean abundance, 0 _10 -5 10 15 20 time. h

Slavov et al. (2014) Cell Rep - proteins

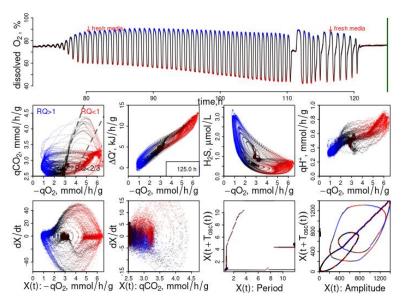
Küenzi, Meyenburg, Fiechter, Sonnleitner at ETH Zürich

.. which didn't matter for the Copenhagen School:

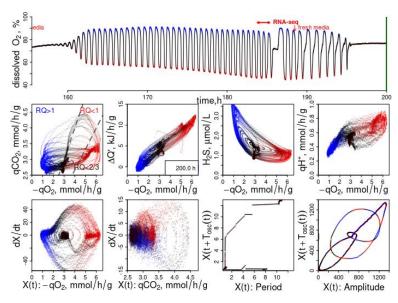
- An intersecting path through the DAG of scientific progress, incl. Monod → Maaløe, Warner, Travers, Gourse, Terry Hwa, Teusink/Bruggeman, Andrea Weisse, the EPCP forum, . . .
- Growth laws: ribosomes as the auto-catalytic core of life, derivation of growth behaviour and optimal gene regulation from first principles.
 Lectures V-VII



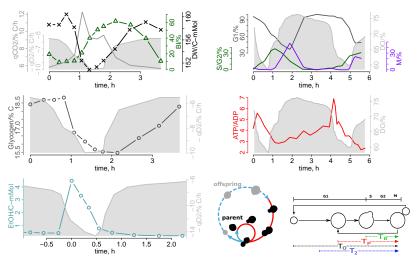
Experiment by Dougie Murray, Jan 2013, Music by Die Goldenen Zitronen.



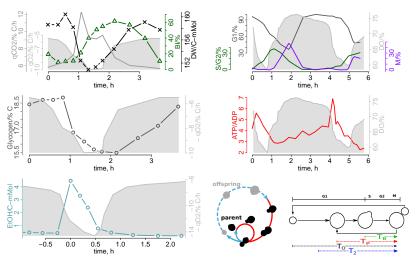
Experiment by Dougie Murray, Jan 2013, Music by Tocotronic.



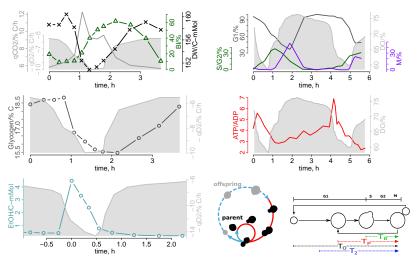
Experiment by Dougie Murray, Jan 2013, Music by Superorganism.



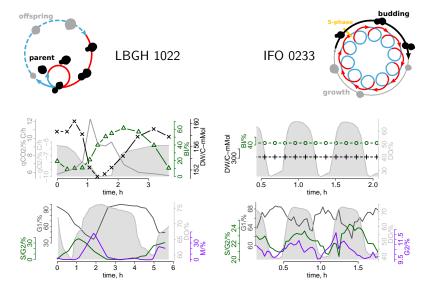
- Fiechter lab (1960s) at ETH Zürich, esp. Kaspar von Meyenburg: the metabolism and energetics of the asymmetric yeast division cycle: budding.
- ▶ Hartwell and Unger (1977): "particularly pertinent prior studies" for clarification of the cell division cycle.

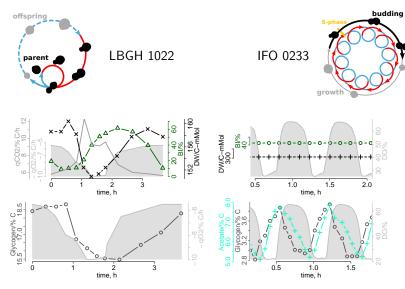


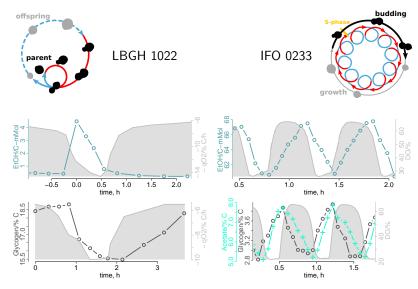
- Budding: S, G2 and M phase; requires glycogen mobilization, and a maximal rate of respiration, with additional overflow metabolism ~ HOC phase.
- ▶ Biomass growth occurs in G1, including build-up of glycogen reserves, at purely respiratory RQ \approx 1 \sim L0C phase.

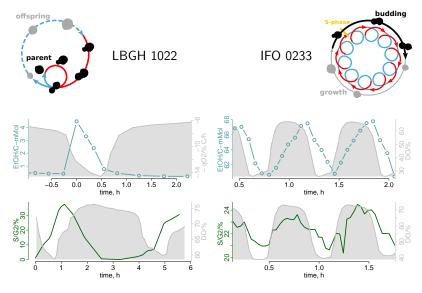


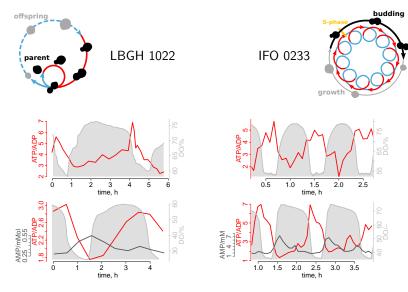
- ▶ The culture medium and conditions established by v. Meyenburg are still used in most studies of respiratory/metabolic oscillations.
- An absurd story of scientific regress occurred with the unfortunate Tu et al. (2005), distorting prior knowledge; we're still recovering.











A Cell Growth Cycle: cell-structural proof-reading?



- nitrogen uptake, AA synthesis,
- translation pulses, high sensitivity to rapamycin (Tor),
- soluble protein peak, autophagy and degradation
- pH, ions, heat dissipation, . . .

Lectures I-IV

buddina

Finding IFO

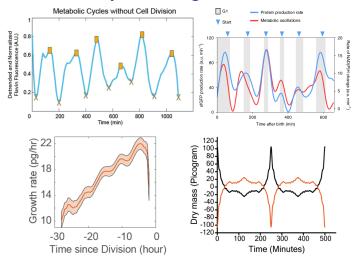


 q_{CO_2} and yield vs. growth rate: **normal!**

- ► Periods and glycogen v growth rate,
- Contraction of phases,
- ► IFO 0233 Rasse II:
 - large lipid inclusions: lipid instead of glycogen?
 - concentric colonies: regular and fast oscillations?

Lectures V-VII

A Cell Growth Cycle in Single Cells

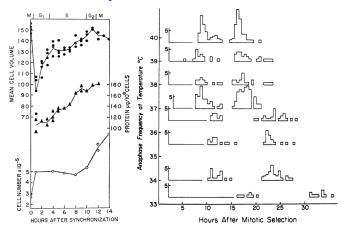


yeast: Cookson et al. (2010), Bryan et al. (2010), Litsios et al. (2019): translation pulses around Start, Baumgartner et al. (2018), Papagiannakis et al. (2017): metabolic oscillations w/o division.

mammalian cells:

Liu et al. (2020), Ghenim et al. (2021): mass density and translation/degradation with $au_{\rm osc}=$ 4–5 h periods.

A Cell Growth Cycle in Mammals?



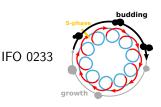
Klevecz and Ruddle (1968), **Science**: Cyclic changes in enzyme activity in synchronized mammalian cell cultures.

Klevecz (1976), PNAS: Quantized generation time in mammalian cells as an expression of the *cellular clock*. \Rightarrow *cell growth cycle* \sim growth rate. Klevecz et al. (2004), PNAS: A genomewide oscillation in *transcription gates* DNA replication and cell cycle. \Rightarrow *pulse-width modulation*.

TODO: Brodsky (1975)

A Cell Growth Cycle in Mammals?





- Another path through the DAG: biological clocks, circadian biology, developmental clocks,
- Bob Klevecz and growth pulses in mammalian cells

Lectures VIII-X

Quantitative Cell Biology

- ► Anabolic vs. catabolic split of Monod model,
 - couple to full metabolic model, FBA to get overall stoichiometries.
- ▶ PWM Model of ribosome production.

References I

Baumgartner, B.L., R. O'Laughlin, M. Jin, L.S. Tsimring, N. Hao, and J. Hasty. 2018. "Flavin-Based Metabolic Cycles Are Integral Features of Growth and Division in Single Yeast Cells." *Sci Rep* 8 (1): 18045.

Behle, A., M. Dietsch, L. Goldschmidt, W. Murugathas, L.C. Berwanger, J. Burmester, L. Yao, et al. 2022. "Manipulation of Topoisomerase Expression Inhibits Cell Division but Not Growth and Reveals a Distinctive Promoter Structure in *Synechocystis*" *Nucleic Acids Res* 50 (22): 12790–12808. https://doi.org/10.1093/nar/gkac1132.

Brodsky, W.Y. 1975. "Protein Synthesis Rhythm." J Theor Biol 55 (1): 167-200.

Bryan, A.K., A. Goranov, A. Amon, and S.R. Manalis. 2010. "Measurement of Mass, Density, and Volume During the Cell Cycle of Yeast." *Proc Natl Acad Sci U S A* 107 (3): 999–1004.

Cookson, N.A., S.W. Cookson, L.S. Tsimring, and J. Hasty. 2010. "Cell Cycle-Dependent Variations in Protein Concentration." *Nucleic Acids Res* 38 (8): 2676–81.

Finn, R.K. 1954. "Accounting for Periodicities in Biology." *Bulletin of Mathematical Biology* 16 (2): 181–82. https://doi.org/10.1007/BF02478374.

Finn, R.K., and R.E. Wilson. 1954. "Population Dynamics of a Continuous Propagator for Microorganisms." *J. Agric. Food Chem.* 2 (2): 66–69. https://doi.org/10.1021/jf60022a609.

Ghenim, L., C. Allier, P. Obeid, L. Herve, J.Y. Fortin, M. Balakirev, and X. Gidrol. 2021. "A New Ultradian Rhythm in Mammalian Cell Dry Mass Observed by Holography." Sci~Rep~11~(1): 1290. https://doi.org/10.1038/s41598-020-79661-9.

Hartwell, L.H., and M.W. Unger. 1977. "Unequal Division in Saccharomyces Cerevisiae and Its Implications for the Control of Cell Division." J Cell Biol 75 (2 Pt 1): 422–35.

Ju, Q., and J.R. Warner. 1994. "Ribosome Synthesis During the Growth Cycle of Saccharomyces Cerevisiae." Yeast 10 (2): 151–57. https://doi.org/10.1002/yea.320100203.

References II

Klevecz, R.R. 1976. "Quantized Generation Time in Mammalian Cells as an Expression of the Cellular Clock." Proc Natl Acad Sci U S A 73 (11): 4012–6.

Klevecz, R.R., J. Bolen, G. Forrest, and D.B. Murray. 2004. "A Genomewide Oscillation in Transcription Gates DNA Replication and Cell Cycle." *Proc Natl Acad Sci U S A* 101 (5): 1200–1205. https://doi.org/10.1073/pnas.0306490101.

Klevecz, R.R., and F.H. Ruddle. 1968. "Cyclic Changes in Enzyme Activity in Synchronized Mammalian Cell Cultures." Science 159 (3815): 634–36.

Libkind, D., C.T. Hittinger, E. Valerio, C. Goncalves, J. Dover, M. Johnston, P. Goncalves, and J.P. Sampaio. 2011. "Microbe Domestication and the Identification of the Wild Genetic Stock of Lager-Brewing Yeast." *Proc Natl Acad Sci U S A* 108 (35): 14539–44. https://doi.org/10.1073/pnas.1105430108.

Lindner, P. 1919. "Das Biosproblem in Der Hefeforschung." Berichte Der Deutschen Botanischen Gesellschaft 37 (11): 34–40. https://doi.org/10.1111/j.1438-8677.1919.tb07801.x.

Litsios, A., D.H.E.W. Huberts, H.M. Terpstra, P. Guerra, A. Schmidt, K. Buczak, A. Papagiannakis, et al. 2019. "Differential Scaling Between G1 Protein Production and Cell Size Dynamics Promotes Commitment to the Cell Division Cycle in Budding Yeast." *Nat Cell Biol* 21 (11): 1382–92. https://doi.org/10.1038/s41556-019-0413-3.

Liu, X., S. Oh, L. Peshkin, and M.W. Kirschner. 2020. "Computationally Enhanced Quantitative Phase Microscopy Reveals Autonomous Oscillations in Mammalian Cell Growth." Proc Natl Acad Sci U S A, October. https://doi.org/10.1073/pnas.2002152117.

Monod, J. 1950. "La Technique de La Culture Continue, Théorie et Applications." *Annales de Institute Pasteur Paris* 79 (4): 390–410.

Novick, A., and L. Szilard. 1950. "Experiments with the Chemostat on Spontaneous Mutations of Bacteria." *Proc Natl Acad Sci U S A* 36 (12): 708–19.

References III

 $\label{eq:papagiannakis} Papagiannakis, A., B. \ Niebel, E.C. \ Wit, and M. \ Heinemann. \ 2017. \ "Autonomous Metabolic Oscillations Robustly Gate the Early and Late Cell Cycle." \ \textit{Mol Cell 65 (2): 285–95. \ https://doi.org/10.1016/j.molcel.2016.11.018.$

Slavov, N., B.A. Budnik, D. Schwab, E.M. Airoldi, and A. van Oudenaarden. 2014. "Constant Growth Rate Can Be Supported by Decreasing Energy Flux and Increasing Aerobic Glycolysis." *Cell Rep* 7 (3): 705–14. https://doi.org/10.1016/j.celrep.2014.03.057.

Tu, B. P., A. Kudlicki, M. Rowicka, and S. L. McKnight. 2005. "Logic of the Yeast Metabolic Cycle: Temporal Compartmentalization of Cellular Processes." *Science* 310 (5751): 1152–8.

Von Meyenburg, H. K. 1968. "Der Sprossungszyklus von Saccharomyces Cerevisiae." Pathobiology 31 (2): 117-27.