The 10kTrees Website: A New Online Resource for Primate Phylogeny

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The comparative method plays a central role in efforts to uncover the adaptive basis for primate behaviors, morphological traits, and cognitive abilities.^{1–4} The comparative method has been used, for example, to infer that living in a larger group selects for a larger neocortex,^{5,6} that primate territoriality favors a longer day range relative to home range size,⁷ and that sperm competition can account for the evolution of primate testes size.^{8,9} Comparison is fundamental for reconstructing behavioral traits in the fossil record, for example, in studies of locomotion and diet.^{10–13} Recent advances in comparative methods require phylogenetic information,^{2,14–16} but our knowledge of phylogenetic information is imperfect. In the face of uncertainty about evolutionary relationships, which phylogeny should one use? Here we provide a new resource for comparative studies of primates that enables users to run comparative analyses on multiple primate phylogenies. Importantly, the 10,000 trees that we provide are not random, but instead use recent systematic methods to create a plausible set of topologies that reflect our certainty about some nodes on the tree and uncertainty about other nodes, given the dataset. The trees also reflect uncertainty about branch lengths.

The comparative method has undergone a revolution in the past 20 years.^{2,14-16} Specifically, new phylogenetic methods provide a way to incorporate evolutionary history directly into comparative research. Phylogeny is essential to comparative research because related species tend to resemble one another, resulting in nonindependent data points.^{2,17,18} Phylogenetic comparative methods can be used to investigate whether two traits change in tandem through time, while also providing the historical scaffolding to identify independent evolutionary origins of the traits

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© 2010 Wiley-Liss, Inc. DOI 10.1002/evan.20251 Published online in Wiley InterScience (www.interscience.wiley.com). of interest. More recently, phylogenetic methods have provided a toolkit to investigate the tempo and mode of evolution,^{19,20} quantify phylogenetic signal in comparative data,^{21,22} and study the factors that influence diversification rates.^{23,24} Computer simulations have revealed that it is usually preferable to conduct comparative tests with some form of phylogenetic method because this reduces false positives (Type I errors) and increases statistical power.17,18,25,26 This latter point is often underappreciated, but it is a logical outcome of phylogenetic comparative analyses that reduce error associated with the estimation of statistics and thus enhance the probability of detecting real effects.26

Researchers generally want to include as many species as possible in a comparative analysis. To incorporate phylogeny in comparative studies of primates, previous researchers have used either published primatewide "supertrees" such as the Purvis phylogeny²⁷ or compiled smaller trees from the literature, often patching these together from among existing phylogenies based on morphol-ogy or genetics.^{28,29} More recently, Bininda-Emonds and coworkers^{30,31} produced a new supertree of mammals. Researchers have begun to use the primate portion of this tree in comparative studies of primates.32-34

The actual tree topology and timing of speciation events is, however, never known with certainty. In addition, phylogenetic relationships should be continually reassessed as new data become available, which recommends against the continued use of older phylogenies such as Purvis',²⁷ since better data are now available. Furthermore,

Box 1. Schematic of Bayesian phylogenetics.

Bayesian methods in phylogenetics typically use Metropoliscoupled Markov chain Monte Carlo algorithms (MCMCMC, or MC³) to generate posterior probability distributions for a set of parameters; that is, the conditional distribution of the parameter given the data. Note that in Bayesian statistics probability cannot be interpreted in its original meaning; rather, it is used to represent parameter uncertainty. In Bayesian phylogenetics, the parameters are comprised of a phylogenetic tree and a specific model of evolution, which is based on the individual priors for these parameters (a priori knowledge or beliefs about a parameter distribution) and the likelihood of the data.

Bayesian MC³ methods start with a random tree and arbitrary initial values for branch lengths and model parameters. In each generation, either a new tree or a new model parameter is proposed. Typically, each proposed change is small; thus, the samples are not random because they are based on the parameter value of the previous generation. The proposal can then be accepted or rejected, depending on the ratio of the posterior densities of the new state to the old state (R). If R>1, an "uphill" step, indicating a state with higher posterior probability, the proposed change is always accepted. If R<1, a "downhill" step, the change is accepted with probability R. The smaller R, the smaller is the proposal acceptance probability. Thus, after a particular number of generations, the region of the parameter space with the highest posterior probability is reached and sampled most often. If the parame-



ter space has multiple peaks that are separated by deep valleys, however, the algorithm may become stuck on a local peak. To rectify that problem, additional chains (socalled hot or heated chains) are used in MC^3 that independently search the tree space, more readily accept proposals with a small R value, and regularly swap states with the original chain (cold chain). Heated chains flatten the posterior probability distribution and thus have shallower valleys, which allows them to more easily cross those valleys.

During MC³, a "chain" of trees is produced that reflects the accepted modifications in phylogenetic information and model parameters. At the beginning of the chain, the likelihood typically climbs quickly, which is called burn-in (see Figure) until the chain eventually reaches its equilibrium distribution (indicated in the Figure by the putative plateau). Every k generations, the cold chain is sampled, which simply entails saving the tree topology,

branch lengths, and all model parameters, and further analyses generally make use of post-burn-in samples. If the posterior probability distribution for each parameter has been approximated adequately, the post-burn-in sampling reflects the true parameter uncertainty and the quality of the sample generally improves as a function of the number of steps (generations). The sample can then be summarized using various statistics, such as histograms, means, or credible intervals. Topology and branch lengths can also be summarized by constructing a majority rule consensus tree with support values in the form of clade credibility values and mean branch lengths. One can also use the whole sample of trees ("tree block") to incorporate topological and branch length uncertainty into comparative analyses that make use of the phylogenetic information. This approach is advocated here and it is also recommended by others.39,40

when conducting a comparative test, it is desirable to incorporate the current level of uncertainty for specific nodes and branch lengths. Indeed, different trees can produce different results in a comparative analysis, making it unwise to condition comparative analyses on a single hypothesis of evolutionary relationships when that hypothesis is legitimately uncertain.³⁵

Some evolutionary anthropologists have accounted for phylogenetic uncertainty by conducting multiple analyses using more than one tree.^{28,36–38} But this raises an important question: How should we decide on the trees to use? A number of authors have proposed that Bayesian phylogenetic approaches provide a way to systematically incorporate phylogenetic uncertainty into comparative research.^{35,39,40} In particular, Bayesian methods allow the user to obtain a set of trees that are sampled in proportion to their posterior probability (Box 1). The set of trees obtained reflects uncertainty in the phylogeny, given the substitution model and data; more certain nodes are found across a greater proportion of the sample of trees, while less certain nodes are found less often. Users can create as many trees as they wish - hundreds, even thousands of phylogenies, all fully bifurcating and with branch lengths, and not simply random permutations of the species in the study.^{41,42} It is possible then to run comparative analyses on this sample and in this way, the results of a comparative study are not conditioned on a particular phylogeny or set of branch lengths.⁴⁰

Here we describe a new online resource for comparative studies of primates, which we call 10kTrees and make available at http://10kTrees.fas. harvard.edu. The 10kTrees website provides a way for users to download up to 10,000 primate phylogenies with branch lengths obtained from a Bayesian phylogenetic analysis. The trees in Version 1 include 189 primate species that are commonly used in comparative research on primates. The data are provided in a standard format⁴³ that can be read by a wide variety of comparative methods programs,^{44,45} including phylogenetics packages for $R.^{46}$ We request that people using this resource cite the current paper, which describes the 10kTrees website and Version 1 of the trees. We are preparing another paper that applies Version 1 to study primate diversification in relation to body mass.⁴⁷

A BAYESIAN INFERENCE OF PRIMATE PHYLOGENY

Bayesian phylogenetic methods provide a way to sample a set of trees in proportion to their posterior probabilities using algorithms based on Metropolis-coupled Markov chain Monte Carlo (MCMCMC, or MC³, see Box 1).^{39,48–50} Nodes that the data strongly support are identical or nearly so across most of the "tree block"; that is, the sample of trees obtained from the analysis. On the consensus tree of the *10kTrees* block, these nodes are thus depicted with high clade credibility values. Some nodes are not well supported, which indicates that alternative arrangements produce similar likelihoods. These nodes vary across the tree block in proportion to their posterior probabilities. By running comparative analyses across this set of trees rather than using a single tree, the results are no longer conditioned on a single tree being correct.

We expect this resource to be especially important for primate phylogenetic comparative studies because it provides a statistically rigorous and principled way to control for uncertainty at various nodes in primate phylogeny.^{51–55} Importantly, our goal for this project is not to produce the definitive primate phylogeny; that goal will be best achieved with more focused studies of gene insertions, whole genomes, and standardized data collection, and will involve longer-term concerted effort by experts in primate phylogenetics (for example, Disotell⁵⁶). Instead, the goal is to produce a set of phylogenetic trees from available data that is appropriate for comparative research on primates. We will, however, regularly update the dataset to accommodate the ever-increasing availability of sequence data and advances in tree inference methods.

For Version 1, we collected data on four mitochondrial genes and one autosomal gene from GenBank. To create the multiple sequence alignments (MSA), we used Muscle 3.7 the default parameters.57 with Because alignment quality can have a substantial impact on the inferred tree,^{58–62} we manually excluded poorly aligned sites or sites with a high percentage of missing data (especially at the beginning and end of the MSA). We constrained 29 major nodes if they were well characterized by at least three genomic Alu insertions.63-69 These constraints eliminate uncertainty at constrained nodes, which we think is reasonable because Alu events are insertion generally regarded as more reliable cladistic indicators that are less prone to homoplasy than are DNA sequence data.63-65 However, we are likely to relax these constraints in future versions of 10kTrees that use additional autosomal loci. Thus, users should

refer to the website for details appropriate for the version they use.

For tree inference in Version 1, we used the program MrBayes 3.1.2.48 Galeopterus variegates, the Sunda flying lemur, or colugo, was identified as the outgroup, since it has been shown that colugos are the closest living relatives to the order Primates.⁷⁰ We ran a Bayesian analysis with two runs and eight chains (one cold chain and seven heated chains) in each run. We used a GTR+I+G substitution model for each of the five genes in a partitioned dataset, which was identified as the best substitution model in the program Find-Model.⁷¹ The analysis for Version 1 was run for eight million generations, with trees sampled every 1,000 generations. We assessed the heating (changed to 0.02) and excluded the first three million generations as burn-in (Box 1). We summarized these topologies by constructing a 50% majority rule consensus tree, which we provide in various graphical formats on the 10kTrees website. Branch lengths were calculated as the mean branch length from all trees in the posterior distribution in which the branch was present. In future versions of 10kTrees, we will provide both molecular branch lengths and, by using fossil calibration points, branches that reflect the time since two species last shared a common ancestor.

APPLICATIONS TO PRIMATE COMPARATIVE BIOLOGY

An important goal of our project is to make the trees readily available for comparative research. To that end, users can download trees in NEXUS⁴³ format. On the 10kTrees website, users can select the number of trees to download. These are sampled from the tree block so that they cover a full range of variation in the analysis; that is, they are sampled evenly along the stored chain of trees rather than simply taking the first n trees in the sample. where n is the number of trees requested by the user. The consensus tree of the full sample is also available to download. In addition, the

user has the option of selecting specific species of interest; the trees are then pruned to the selected species before download. In terms of the actual data used to generate the trees, users can obtain the original sequence data, the list of species that were studied, an availability matrix for the distribution of genetic data across species, and details on how the data were analyzed (that is, the substitution model and parameters of the MC³ analysis, such as sampling rate, number of chains, and number of sampled generations). The phylogenetic constraints based on Alu insertions are also downloadable from the website. By providing the raw data files, users can easily rerun the phylogenetic analysis in a different computer package or with different settings, e.g., without the constraints.

The phylogenies available from 10kTrees provide a significant improvement over the two primatewide phylogenies that are currently most commonly used in comparative research.^{27,30} The older of these phylogenies, published by Andy Purvis in 1995,²⁷ was a ground-breaking contribution to comparative primatology when it was published. However, fewer genetic data were available in 1995, and many polytomies were present in the tree. In particular, the Asian colobines showed almost no resolution because phylogenetic information for this clade was generally unavailable when the tree was constructed. In addition to its lack of resolution, the Purvis phylogeny includes a number of topological misplacements even at the generic level as assessed by comparison to the preponderance of DNA sequence and Alu insertion data used to infer the 10kTrees. For example, the basal bifurcation of the platyrrhines is incorrect on the basis of Alu insertions, as are the positions of Aotus and Callicebus.65 Recent sequence-based studies and our research clearly contradict the topology given for other genera such as *Callimico* and *Lophocebus*.^{72,73} Given its lack of resolution and topological misplacements at the generic level, continued use of the Purvis phylogeny²⁷ cannot be recommended.

The primate portion of the more recent Bininda-Emonds supertree³⁰ is an improvement over Purvis' phylogeny,²⁷ but it still suffers from excessive polytomies within the guenons and the Asian colobines. The Bininda-Emonds tree contains an incorrect topology for the basal bifurcations among the platyrrhines, given the Alu insertion data and whole mitochondrial genome evidence.65,72 It also includes apparent anomalies or errors, such as the paraphyletic placement of *Callicebus per*sonatus as separate from its congeners and basal to Cebidae. While the Bininda-Emonds tree is useful for mammal-wide comparative analyses, we recommend 10kTrees for studies focused on primates.

We designed the website so that it can be easily updated as new genetic data become available to infer primate phylogeny. Thus, the website provide multiple versions, will including an "archive" where previous versions can be accessed. We are already working on Version 2 of the dataset, which will include over 230 species and more genes. We also expect that the website itself will evolve to provide more tools for primate comparative biology. In future versions, for example, we plan to provide a taxonomic translation tool. Thus, readers will be able to select species based on their names from GenBank, or from lists of names in which the original species designations are translated to commonly used taxonomies, such as the taxonomies by Corbet and Hill⁷⁴ and Groves in Wilson and Reeder.75 In addition to giving trees with branch lengths proportional to genetic change, a future version of the website will provide dated trees based on fossil calibration points. We will also make available scripts to link the trees to comparative data to produce a single NEXUS⁴³ file for analysis. We will provide advice for importing the trees into analysis programs, such as BayesTraits,⁴⁴ R,⁷⁶ and Mesquite.⁴⁵ Ultimately, we aim for a flexible, open-access, and user-friendly platform that will enhance the use of phylogenetic approaches in primate evolution, and will grow as new sequence data become available.

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REFERENCES

1 Clutton-Brock TH, Harvey PH. 1977. Primate ecology and social organization. J Zool 183:1–39.

2 Nunn CL, Barton RA. 2001. Comparative methods for studying primate adaptation and allometry. Evol Anthropol 10:81–98.

3 Martin RD. 1990. Primate origins and evolution. London: Chapman and Hall.

4 Plavcan JM, van Schaik CP. 1992. Intrasexual competition and canine dimorphism in anthropoid primates. Am J Phys Anthropol 87:461–477.

5 Dunbar RIM. 1998. The social brain hypothesis. Evol Anthropol 6:178–190.

6 Barton RA. 1996. Neocortex size and behavioural ecology in primates. Proc R Soc London, B 263:173–177.

7 Mitani JC, Rodman PS. 1979. Territoriality: the relation of ranging pattern and home range size to defendability, with an analysis of territoriality among primate species. Behav Ecol Sociobiol 5:241–251.

8 Harcourt AH, Harvey PH, Larson SG, Short RV. 1981. Testis weight, body weight and breeding system in primates. Nature 293:55–57.

9 Harcourt AH, Purvis A, Liles L. 1995. Sperm competition: mating system, not breeding season, affects testes size of primates. Funct Ecol 9:468–476.

10 Spoor F, Garland T, Krovitz G, Ryan TM, Silcox MT, Walker A. 2007. The primate semicircular canal system and locomotion. Proc Natl Acad Sci USA 104:10808–10812.

11 Jungers WL, Godfrey LR, Simons EL, Chatrath PS. 1997. Phalangeal curvature and positional behavior in extinct sloth lemurs (Primates, Palaeopropithecidae). Proc Natl Acad Sci USA 94:11998–12001.

12 Napier JR, Walker AC. 1967. Vertical clinging and leaping: a newly recognized category of locomotor behaviour of primates. Folia Primatol 6:204–219.

13 Kay RF. 1984. On the use of anatomical features to infer foraging behavior in extinct primates. In: Rodman P, Cant J, editors. Adaptations for foraging in nonhuman primates. New York: Columbia University Press.

14 Garland T, Harvey PH, Ives AR. 1992. Procedures for the analysis of comparative data using phylogenetically independent contrasts. Syst Biol 4:18–32.

15 Felsenstein J. 1985. Phylogenies and the comparative method. Am Nat 125:1–15.

16 Harvey PH, Pagel MD. 1991. The comparative method in evolutionary biology. Oxford: Oxford University Press.

17 Martins EP, Garland T. 1991. Phylogenetic analyses of the correlated evolution of continuous characters: a simulation study. Evolution 45:534–557.

18 Purvis A, Gittleman JL, Luh H. 1994. Truth or consequences: effects of phylogenetic accu-

racy on two comparative methods. J Theor Biol 167:293–300.

19 Pagel M. 1997. Inferring evolutionary processes from phylogenies. Zool Scripta 26:331–348.

20 Pagel M. 1999. Inferring the historical patterns of biological evolution. Nature 401:877-884.

21 Blomberg SP, Garland T, Ives AR. 2003. Testing for phylogenetic signal in comparative data: behavioral traits are more labile. Evolution 57:717–745.

22 Freckleton RP, Harvey PH, Pagel M. 2002. Phylogenetic analysis and comparative data: a test and review of evidence. Am Nat 160:712–726.

23 Nee S, Holmes EC, May RM, Harvey PH. 1994. Extinction rates can be estimated from molecular phylogenies. Philos Trans R Soc London B 344:77–82.

24 Maddison WP. 2006. Confounding asymmetries in evolutionary diversification and character change. Evolution 60:1743–1746.

25 Symonds MRE. 2002. The effects of topological inaccuracy in evolutionary trees on the phylogenetic comparative method of independent contrasts. Syst Biol 51:541–553.

26 Rohlf FJ. 2006. A comment on phylogenetic correction. Evolution 60:1509–1515.

27 Purvis A. 1995. A composite estimate of primate phylogeny. Philos Trans R Soc London, Series B 348:405–421.

28 Mitani JC, GrosLouis J, Manson JH. 1996. Number of males in primate groups: Comparative tests of competing hypotheses. Am J Primatol 38:315–332.

29 Smith RJ, Cheverud JM. 2002. Scaling of sexual dimorphism in body mass: a phylogenetic analysis of Rensch's rule in primates. Int J Primatol 23:1095–1135.

30 Bininda-Emonds ORP, Cardillo M, Jones KE, MacPhee RDE, Beck RMD, Grenyer R, Price SA, Vos RA, Gittleman JL, Purvis A. 2007. The delayed rise of present-day mammals. Nature 446:507–512.

31 Bininda-Emonds ORP, Cardillo M, Jones KE, MacPhee RDE, Beck RMD, Grenyer R, Price SA, Vos RA, Gittleman JL, Purvis A. n.d. Corrigendum: the delayed rise of present-day mammals. Nature. In press.

32 Davies TJ, Pedersen AB. 2008. Phylogeny and geography predict pathogen community similarity in wild primates and humans. Proc R Soc B 275:1695–1701.

33 Walker RS, Gurven M, Burger O, Hamilton MJ. 2008. The trade-off between number and size of offspring in humans and other primates. Proc R Soc B 275:827–833.

34 Nunn CL, McNamara P, Capellini I, Preston P, Barton RA. 2009. Primate sleep in phylogenetic perspective. In: McNamara P, Barton RA, Nunn CL, editors. Evolution of sleep: phylogenetic and functional perspectives. Cambridge: Cambridge University Press.

35 Lutzoni F, Pagel M, Reeb V. 2001. Major fungal lineages are derived from lichen symbiotic ancestors. Nature 411:937–940.

36 Kamilar J, Martin S, Tosi A. 2009. Combining biogeographic and phylogenetic data to examine primate speciation: an example using cercopithecin monkeys. Biotropica 41:514–519.

37 Thierry B, Iwaniuk A, Pellis S. 2000. The influence of phylogeny on the social behaviour of macaques (Primates: Cercopithecidae, genus Macaca). Ethology 106:713–728.

38 Nunn CL. 1999. The number of males in primate social groups: a comparative test of the socioecological model. Behav Ecol Sociobiol 46:1–13.

39 Huelsenbeck JP, Rannala B, Masly JP. 2000. Accommodating phylogenetic uncertainty in evolutionary studies. Science 288:2349–2350.

40 Pagel M, Lutzoni F. 2002. Accounting for phylogenetic uncertainty in comparative studies of evolution and adaptation. In: Lässig M, Valleriani A, editors. Biological evolution and statistical physics. Berlin: Springer-Verlag. p 148–161.

41 Martins EP. 1996. Conducting phylogenetic comparative studies when the phylogeny is not known. Evolution 50:12–22.

42 Abouheif E. 1998. Random trees and the comparative method: a cautionary tale. Evolution 52:1197–1204.

43 Maddison DR, Swofford DL, Maddison WP. 1997. Nexus: an extensible file format for systematic information. Syst Biol 46:590–621.

44 Pagel M, Meade A. 2007. Bayestraits (www. evolution.rdg.ac.uk). Version 1.0. Reading, UK.

45 Maddison WP, Maddison DR. 2006. Mesquite: a modular system for evolutionary analysis. Version 2.5: http://mesquiteproject.org.

46 R Development Core Team. 2009. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL http://www.R-project.org.

47 Matthews LJ, Arnold C, Machanda Z, Nunn C. n.d. Body size effects on speciation and extinction rates in primates. In preparation.

48 Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19:1572–1574.

49 Larget B, Simon DL. 1999. Markov Chasin Monte Carlo algorithms for the Bayesian analysis of phylogenetic trees. Mol Biol Evol 16:750–759.

50 Yang Z, Rannala B. 1997. Bayesian phylogenetic inference using DNA sequences: a Markov Chain Monte Carlo method. Mol Biol Evol 14:717–724.

51 Martin RD. 2008. Evolution of placentation in primates: implications of mammalian phylogeny. Evol Biol 35:125–145.

52 Ting N, Tosi AJ, Li Y, Zhang YP, Disotell TR. 2008. Phylogenetic incongruence between nuclear and mitochondrial markers in the Asian colobines and the evolution of the langurs and leaf monkeys. Mol Phylogenet Evol 46:466–474.

53 Finarelli JA, Clyde WC. 2004. Reassessing hominoid phylogeny: evaluating congruence in the morphological and temporal data. Paleobiology 30:614–651.

54 Schneider H. 2000. The current status of the New World monkey phylogeny. Anais Acad Brasileira Ciencias 72:165–172.

55 Schmitz J, Roos C, Zischler H. 2005. Primate phylogeny: molecular evidence from retroposons. Cytogenet Genome Res 108:26–37.

56 Disotell TR. 2008. Primate phylogenetics: encyclopedia of life sciences. Chinchester: John Wiley and Sons.

57 Edgar RC. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res 32:1792.

58 Ogden TH, Rosenberg MS. 2006. Multiple sequence alignment accuracy and phylogenetic inference. Syst Biol 55:314–328.

59 Talavera G, Castresana J. 2007. Improvement of phylogenies after removing divergent

and ambiguously aligned blocks from protein sequence alignments. Syst Biol 56:564–577.

60 Smythe AB, Sanderson MJ, Nadler SA. 2006. Nematode small subunit phylogeny correlates with alignment parameters. Syst Biol 55:972– 992.

61 Kjer KM. 1995. Use of rRNA secondary structure in phylogenetic studies to identify homologous positions: an example of alignment and data presentation from the frogs. Mol Phylogenet Evol 4:314–330.

62 Morrison DA, Ellis JT. 1997. Effects of nucleotide sequence alignment on phylogeny estimation: a case study of 18S rDNAs of Apicomplexa. Mol Biol Evol 14:428–441.

63 Ray DA, Batzer MA. 2005. Tracking Alu evolution in New World primates. BMC Evol Biol 5:51.

64 Ray DA, Xing JC, Hedges DJ, Hall MA, Laborde ME, Anders BA, White BR, Stoilova N, Fowlkes JD, Landry KE. 2005. Alu insertion loci and platyrrhine primate phylogeny. Mol Phylogenet Evol 35:117–126.

65 Xing JC, Witherspoon DJ, Ray DA, Batzer MA, Jorde LB. 2007. Mobile DNA elements in primate and human evolution. Am J Phys Anthropol suppl. 45:2–19.

66 Xing J, Wang H, Han KD, Ray DA, Huang CH, Chemnick LG, Stewart CB, Disotell TR, Ryder OA, Batzer MA. 2005. A mobile element based phylogeny of Old World monkeys. Mol Phylogenet Evol 37:872–880.

67 Schmitz J, Ohme M, Zischler H. 2001. SINE insertions in cladistic analyses and the phylogenetic affiliations of *Tarsius bancanus* to other primates. Genetics 157:777–784.

68 Salem AH, Ray DA, Xing J, Callinan PA, Myers JS, Hedges DJ, Garber RK, Witherspoon DJ, Jorde LB, Batzer MA. 2003. Alu elements and hominid phylogenetics. Proc Nat Acad Sci USA 100:12787–12791.

69 Roos C, Schmitz J, Zischler H. 2004. Primate jumping genes elucidate strepsirrhine phylogeny. Proc Nat Acad Sci USA 101:10650–10654.

70 Janecka JE, Miller W, Pringle TH, Wiens F, Zitzmann A, Helgen KM, Springer MS, Murphy WJ. 2007. Molecular and genomic data identify the closest living relative of primates. Science 318:792–794.

71 Tao N, Richardson R, Bruno W, Kuiken C. 2005. FindModel (http://hcv.lanl.gov/content/hcv-db/findmodel/findmodel.html).

72 Hodgson JA, Sterner KN, Matthews LJ, Burrell AS, Jani RA, Raaum RL, Stewart CB, Disotell TR. 2009. Successive radiations, not stasis, in the South American primate fauna. Proc Nat Acad Sci USA 106:5534–5539.

73 Burrell A, Jolly C, Tosi A, Disotell T. 2009. Mitochondrial evidence for the hybrid origin of the kipunji, *Rungweebus kipunji* (Primates: Papionini). Mol Phylogenet Evol 51:340– 348.

74 Corbet GB, Hill JE. 1991. A world list of mammalian species. Oxford: Oxford University Press.

75 Wilson DE, Reeder DM. 2005. Mammal species of the world: Johns Hopkins University Press.

76 R Development Core Team. 2008. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3–900051-07–0, URL http://www.R-project.org.

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