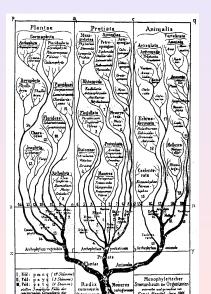
Lecture: Some Topics in Phylogenetics

Lecturer: Dr. Stephan Steigele

Bioinf, University of Leipzig

Leipzig WS07/08

Phylogeny



Part I

Introduction to Phylogenetics

Outline

Source Materials Introduction to Evolutionary Analysis What is Phylogenetic Analysis? **Molecular Phylogenetics** Introduction to Phylogenetic Trees What are Phylogenetic Trees? **Enumerating Trees** Computer Representation of a phylogenetic tree Nested Structure Printing a phylogenetic tree Algorithm for printing a phylogenetic tree Parsing a phylogenetic tree Drawing a phylogenetic tree Computing a circular embedding Computing the edge angles Determining coordinates Example

-Source Materials

Main sources for this lecture:

Some parts of this lecture were simply taken from scripts developed at University Tuebingen from **Prof. Daniel Huson** and in part by **Dr. Kay Nieselt** between 2002 and 2005. **More important**: All parts of the lecture are highly inspired by the following book:

 Jospeh Felsenstein, Inferring Phylogenetics, Sinauer Associates, 2004

Reading of the book is highly recommended ..

-Source Materials

Additional Material

More sources for this lecture:

- R. Durbin, S. Eddy, A. Krogh & G. Mitchison, Biological sequence analysis, Cambridge, 1998
- J. Setubal & J. Meidanis, Introduction to computational molecular biology, 1997.
- D.W. Mount. Bioinformatics: Sequences and Genome analysis, 2001.
- D.L. Swofford, G.J. Olsen, P.J.Waddell & D.M. Hillis, Phylogenetic Inference, in: D.M. Hillis (ed.), Molecular Systematics, 2 ed., Sunderland Mass., 1996.

- -Introduction to Evolutionary Analysis
 - What is Phylogenetic Analysis?

Phylogenetic analysis

Given a collection of extant species. The goal of phylogenetic analysis is to determine and describe the **evolutionary relationship** between the species. In particular, this involves determining the order of speciation events and their approximate timing.

It is generally assumed that **speciation** is a branching process: a population of organisms becomes separated into two sub-populations. Over time, these evolve into separate species that do not cross-breed.

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 - What is Phylogenetic Analysis?

Phylogenetic analysis

Because of this assumption, a **tree** is often used to represent a proposed phylogeny for a set of species, showing how the species evolved from a common ancestor. We will study this in detail.

However, there are a number of situations in which a tree is less appropriate than a phylogenetic **network**. We will study this in a later Chapter.

- -Introduction to Evolutionary Analysis
 - Molecular Phylogenetics

Definitions of Molecular Phylogenetics

- Molecular phylogenetics refers to any method of inferring evolutionary relationships from similarities or differences in molecular structure.
- However: Molecular characters suffer from problems that also afflict morphological characters.
- For example, neither molecules nor morphology may be able to resolve the phylogeny of evolution that was both ancient and rapid, as in the Cambrian Explosion.

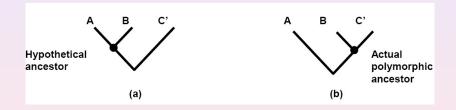
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The Character of evolution

- One problem shared by molecular and morphological characters is **homoplasy** (nonhomologous characters appearing to be similar in different taxa).
- Another problems shared by molecular and morphological phylogenetics arise from **polymorphism** (homologous characters appearing differently in the same species). Because of polymorphism, the time of divergence may appear to be earlier than it was.
- Polymorphism can also result in the incorrect phylogenetic sequence.
- Similar problems result from different copies of duplicated genes

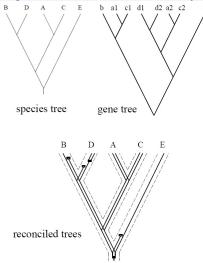
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Example: Polymorphism



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Example: Gene Tree Vs Species Tree



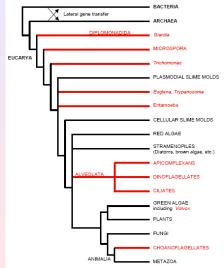
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Some Problems and new insights

- Another problem with molecular phylogenetics is long-branch attraction: the tendency of fast-evolving molecules to appear more closely related than they actually are.
- Molecular phylogenetics has gained wide acceptance in spite of these and other problems because it provides a large amount of evidence that is independent of morphology, as well as other advantages.
- Several kinds of experiments support the validity of molecular phylogenetics
- Because of long-branch attraction, differences in sequence alignment, limitations in the size of study groups, and different methods of tree reconstruction, conflicting molecular phylogenies have been proposed.

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Example: Polyphyletic Origin of Protozoans



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Molecular Characters

- Molecular characters can be of two types: discrete (qualitative) differences in molecular sequence and continuous (quantitative) distance between molecules.
- The first step in molecular phylogenetics is to select a suitable molecule that is **homologous** in all the taxa to be included in the phylogeny.
- Many molecular characters are much less susceptible to homoplasy and longbranch attraction than are nucleic-acid sequences.
- These characters include amino-acid sequences, the positions of short and long interspersed elements, and Hox genes.

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Molecular Characters

- Elongation factors, actin, and tubulins are among the widely used proteins.
- The positions of short and long interspersed elements (SINEs and LINEs) are another increasingly common source of discrete characters.
- Hox genes have also been used to infer phylogenetic relationships.
- The most commonly used molecular data for higher taxonomic levels are base sequences from genes that encode ribosomal RNA, especially 18S rDNA.

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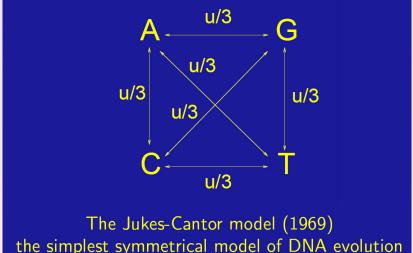
Modeling Evolution

- Assumptions may be needed about the probabilities of different molecular changes.
- Molecular relationships are represented as trees constructed of branches with nodes at both ends of each branch.
- Inferring (reconstructing) a phylogeny consists of creating or selecting one tree out of perhaps millions of possible ones.

Introduction to Evolutionary Analysis

Molecular Phylogenetics

Jukes-Cantor Model



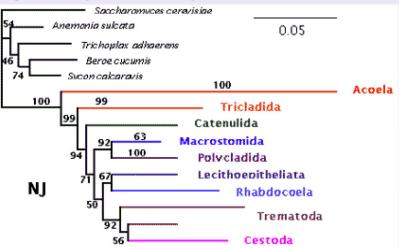
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Many Ways to do it !!

- The neighbor-joining method (distance based) is an algorithm that generates one tree with the shortest total branch length.
- The maximum parsimony method (character based) selects the cladogram with the minimum number of changes in character state.
- The maximum likelihood method (character based) begins with an explicit model of evolution and possible trees, then it attempts to find the tree that is most likely with the given data.
- With more than a few taxa, any method requires a computer.
- Phylogenies reconstructed by different methods are generally similar to each other

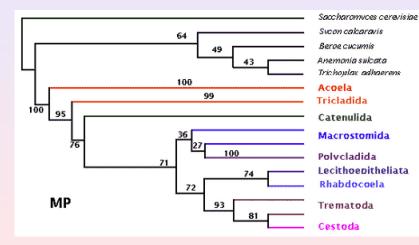
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Example: Neighbor-Joining Tree

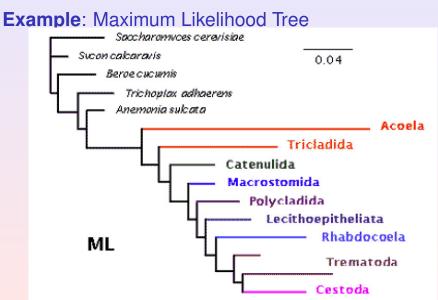


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Example: Maximum Parsimony Tree



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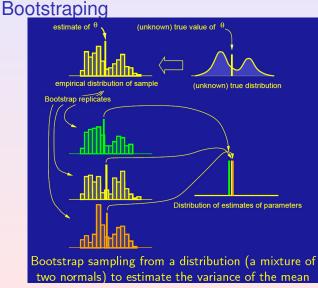


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Getting Confidence

- Confidence in an internal branch can be tested by bootstrapping
- A branch with low bootstrap support may be collapsed.
- Molecular and morphological data can be combined to create a total-evidence tree.

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Collapsed Tree

A **consensus tree** can be created by collapsing branches that are not supported in all trees created by different methods of analysis. A consensus tree can also be produced by comparing molecular and morphological trees.

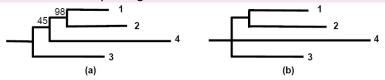


Figure 13. Collapsing branches that are poorly supported. (a) The original tree with one branch having a bootstrap value of only 45%. (b) After collapsing the poorly supported branch there is an unresolved trichotomy for branches (1 + 2), 3, and 4.

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Consistency

- As techniques have improved and more molecules from more species have been sequenced, many of the past conflicts have been resolved.
- Testing the Validity of Traditional Morphological Characters by Seeing Whether They Are Consistent With Molecular Trees
- To be consistent with a given phylogenetic tree, a character must map onto the tree with few changes in character state.

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Examples for Consistency

For example, bilateral symmetry is consistent with the traditional morphology based cladogram for the Big Nine phyla (those with more than 5,000 named species), since it requires only one change in character state.

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Example: Bilateral Symmetry

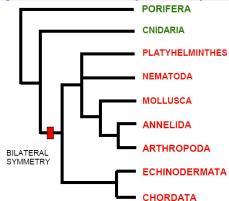


Figure 14. The traditional morphology-based phylogeny of the major animal phyla, based on the "hypothetical diagram" by Hyman (1940, vol. 1, p. 38). Bilateral symmetry is consistent with this phylogeny because it requires only one change in character state to account for its distribution among the phyla.

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Examples for Consistency

Segmentation, however, is less consistent with traditional morphology based phylogenetic trees, because it requires at least two changes in character state: one for annelids and arthropods and one for chordates.

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Example: Segmentation

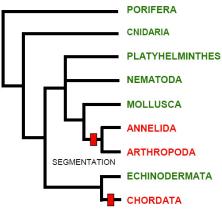


Figure 15. Traditional phylogeny of the Big Nine showing that segmentation is less consistent, because it requires at least two changes in character state to account for its distribution.

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Examples for Consistency

Lack of consistency implies that either the character is not synapomorphic (homologous), or the phylogenetic tree is incorrect.

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Combining Evidence

- A character that is consistent with both a morphological and a molecular phylogeny is more likely to be phylogenetically informative
- Morphological characters have not led to a consensus phylogeny.
- The following morphological characters traditionally used in phylogenetics are also consistent with the widely accepted molecular phylogenetic tree of the Big Nine Phyla: bilateral symmetry and triploblasty, deuterostomy, and spiral cleavage pattern.

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Example: Morpholocial Markers that fit Molecular Analysis

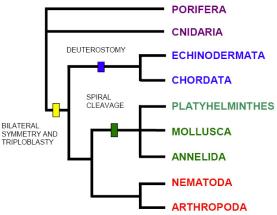


Figure 16. Bilateral symmetry and triploblasty, deuterostomy, and spiral cleavage pattern are

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Combining Evidence

- Bilateral symmetry and triploblasty are also consistent with a molecular phylogenetic tree that includes all animal phyla.
- Deuterostomy in Echinodermata, Hemichordata, and Chordata is also consistent in a molecular phylogenetic tree of all animal phyla.
- The spiral-cleavage pattern is somewhat consistent with a molecular phylogenetic tree that includes all animal phyla.

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How Much Information??

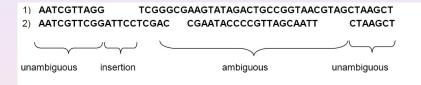


Figure 2. Homologous sequences of DNA bases from two taxa (1 and 2). An insertion, deletions, and a large number of mutations in the middle portion of the sequence make alignment ambiguous for that region.

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Informative Sites in Molecular Data

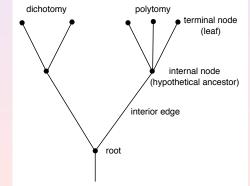
| 1) | т | т | С | G | Α | С | С | G | т | |
|----|---|---|---|---|---|---|---|---|---|--|
| 2) | с | т | т | Α | Α | с | т | G | т | |
| 3) | с | т | Α | т | G | с | т | G | G | |
| 4) | с | т | G | т | G | с | с | G | G | |
| | | | | | x | | у | | z | |

Figure 7. Aligned homologous DNA sequences from four taxa. Informative sites are indicated by letters x, y, and z. Only positions with two or more different bases, at least two of which occur in more than one taxon, are informative.

-What are Phylogenetic Trees?

A small tree nursery

A tree consists of **nodes** connected by **branches** (also called **edges**). Terminal nodes (also called **leaves**, OTUs) represent sequences or species for which we have data. Internal nodes represent hypothetical ancestors. If an internal node has three branches, it has



What are Phylogenetic Trees?

Phylogenetic trees

In the following, we will use $X = \{x_1, x_2, ..., x_n\}$ to denote a set of **taxa**, where a **taxon** x_i is simply a representative of a group of individuals defined in some way.

A **phylogenetic tree** (on *X*) is a system $T = (V, E, \lambda)$ consisting of a connected graph (V, E) without cycles, together with a **labeling** λ of the leaves by elements of *X*, such that:

- 1. every leaf is labeled by exactly one taxon, and
- 2. every taxon appears exactly once as a label.

- What are Phylogenetic Trees?

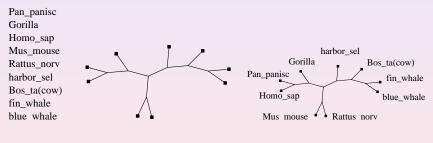
Phylogenetic trees

- ► Further, we require that either all internal nodes have degree ≥ 3, in which case T is called unrooted,
- or there exists precisely one internal root node ρ of degree 2, and T is called rooted.
- A phylogenetic tree *T* is called **binary**, if every internal node *v* has degree 3, except *ρ*, if *T* is rooted.
- An X-tree is obtained by relaxing the definition to allow labels on internal nodes, or nodes to carry multiple labels.
- Occasionally we will allow an arbitrary node to be the root.

-What are Phylogenetic Trees?

Unrooted trees

An unrooted phylogenetic tree is obtained by placing a set of taxa on the leaves of a tree:



Taxa X + tree \Rightarrow

phylogenetic tree T on X

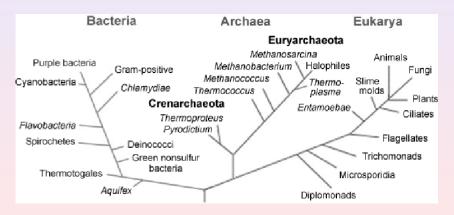
Unrooted trees are often displayed using this type of **circular** layout.

-Introduction to Phylogenetic Trees

-What are Phylogenetic Trees?

Rooted trees

A rooted tree is usually drawn with the root placed at the bottom, top or left of the figure:



Stephan Steigele

Modern version of the tree of life.

- What are Phylogenetic Trees?

Edge lengths

Consider a phylogenetic tree T on X. The **topology** of the tree describes the putative order of speciation events that gave rise to the extant taxa.

Additionally, we can assign **lengths** to the edges of the tree. Ideally, these lengths should represent the amount of time that lies between two speciation events. However, **in practice the edge lengths usually represent quantities obtained by some given computation and only correspond very indirectly to time**.

-Introduction to Phylogenetic Trees

What are Phylogenetic Trees?

Edge lengths

In the following, we will use $\omega : E \to \mathbb{R}_{\geq 0}$ to specify edge lengths and will use $T = (V, E, \lambda, \omega)$ to denote a phylogenetic tree with edge lengths.

Enumerating Trees

The number of edges and nodes of an unrooted phylogenetic tree

Let T be an unrooted phylogenetic tree on n taxa, i.e., with n leaves. How many nodes and edges does T have? Let us assume that T is binary. Any non-binary tree on n taxa will have less nodes and edges.

Lemma

A binary phylogenetic tree T on n taxa has 2n - 2 nodes, 2n - 3 edges and n - 3 interior edges for all $n \ge 3$.

- Enumerating Trees

The number of edges and nodes of an unrooted phylogenetic tree

Proof.

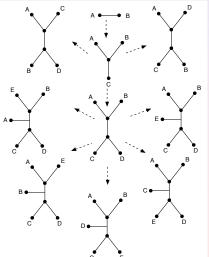
by induction. For n = 3 there is exactly one tree: it has 3 outer edges and zero interior edges. Also it has 3 leaves and one interior node.

We assume that the lemma has been proven for *n*. Any tree *T'* with n + 1 leaves can be obtained from some tree *T* with *n* leaves by inserting a new node *v* into some edge *e* of *T* and connecting *v* to a new leaf *w*. This increases both the number of nodes and the number of edges by 2. Thus *T'* has 2n - 3 + 2 = 2n - 1 = 2(n + 1) - 3 edges. The number of interior edges is (2n - 3) - n = n - 3.

-Introduction to Phylogenetic Trees

- Enumerating Trees

The number of edges and nodes of an unrooted phylogenetic tree



- Enumerating Trees

The number of phylogenetic trees

An unrooted tree T with n leaves has 2n - 2 nodes and 2n - 3edges. A root can be added in any of the 2n - 3 edges, thus producing 2n - 3 different rooted trees from T: h а а a

- Enumerating Trees

Number of phylogenetic trees

For n = 3 there are three ways of adding a root. Similarly, there are 3 different ways of adding an extra edge with a new leaf to obtain an unrooted tree on 4 leaves. This new tree has (2n - 3) = 5 edges and there are 5 ways to obtain a new tree with 5 leaves etc.

Continuing this, we see that there are

$$U(n) = (2n-5)!! := 3 \cdot 5 \cdot 7 \cdot \cdots \cdot (2n-5)$$

unrooted trees on n leaves. Similarly, there are

$$R(n) = (2n-3)!! = U(n) \cdot (2n-3) = 3 \cdot 5 \cdot \dots \cdot (2n-3)$$

rooted trees.

These numbers grows very rapidly with *n*, for example, $U(10) \approx 2$ million and $U(20) \approx 2.2 \times 10^{20}$.

-Introduction to Phylogenetic Trees

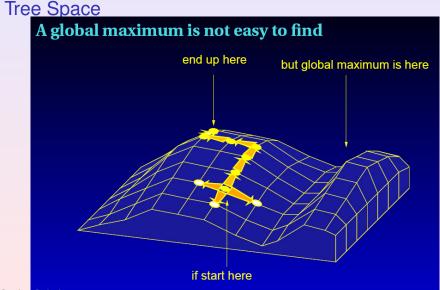
- Enumerating Trees

Counting of Trees

| species | number of trees |
|---------|-------------------------------|
| 1 | 1 |
| 2 | 1 |
| 3 | 3 |
| 4 | 15 |
| 5 | 105 |
| 6 | 945 |
| 7 | 10,395 |
| 8 | 135,135 |
| 9 | 2,027,025 |
| 10 | 34,459,425 |
| 11 | 654,729,075 |
| 12 | 13,749,310,575 |
| 13 | 316,234,143,225 |
| 14 | 7,905,853,580,625 |
| 15 | 213,458,046,676,875 |
| 16 | 6,190,283,353,629,375 |
| 17 | 191,898,783,962,510,625 |
| 18 | 6,332,659,870,762,850,625 |
| 19 | 221,643,095,476,699,771,875 |
| 20 | 8,200,794,532,637,891,559,375 |
| 30 | $4.9518 	imes 10^{38}$ |
| 40 | 1.00985×10^{57} |
| 50 | 2.75292×10^{76} |

-Introduction to Phylogenetic Trees

- Enumerating Trees



- Computer Representation of a phylogenetic tree

Computer representation of a phylogenetic tree

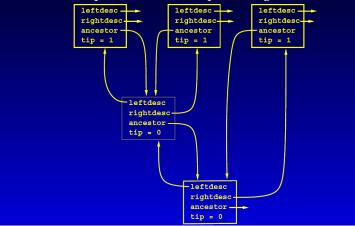
Let X be a set of taxa and T a phylogenetic tree on X. To represent a phylogenetic tree in a computer we maintain a set of nodes V and a set of edges E.

- ► Each edge e ∈ E maintains a reference to its source node s(e) and target node t(e).
- ► Each node v ∈ V maintains a list of references to all incident edges.
- ► Each node v ∈ V maintains a reference λ(v) to the taxon that it is labeled by, and, vice versa, ν(x) maps a taxon x to the node that it labels.
- Each edge $e \in E$ maintains its length $\omega(e)$.

- Computer Representation of a phylogenetic tree

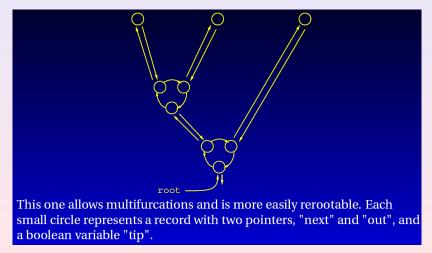
Computer Representation

Using records (in C: structures, in Java and C++: classes) and pointers: Here is one record-pointer structure representing a small tree:



- Computer Representation of a phylogenetic tree

Computer Representation allowing Multifurcations



- Computer Representation of a phylogenetic tree

-Nested Structure

Nested structure

A rooted phylogenetic tree $T = (V, E, \lambda)$ is a **nested** structure:

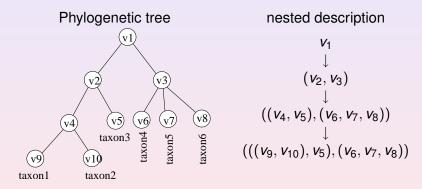
- Consider any node $v \in V$.
- Let T_v denote the subtree rooted at v.
- Let v_1, v_2, \ldots, v_k denote the children of v.
- ► Then T_v is obtainable from its subtrees T_{v1}, T_{v2},..., T_{vk} by connecting v to each of their roots.



- Computer Representation of a phylogenetic tree

-Nested Structure

Such a nested structure can be written using nested brackets:



Description: (((*taxon*₁, *taxon*₂), *taxon*₃), (*taxon*₄, *taxon*₅, *taxon*₆))

- Computer Representation of a phylogenetic tree

Printing a phylogenetic tree

Printing a phylogenetic tree

Phylogenetic trees are usually printed using the so-called **Newick** format. The Newick Standard for representing trees in computer-readable form makes use of the correspondence between trees and nested parentheses, noticed in 1857 by the famous English mathematician Arthur Cayley.

- Computer Representation of a phylogenetic tree

-Printing a phylogenetic tree

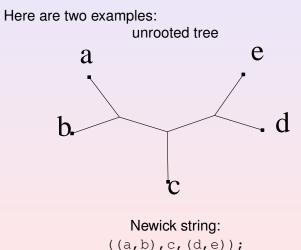
Newick Format

- The tree ends with a semicolon.
- Interior nodes are represented by a pair of matched parentheses.
- Between them are representations of the nodes that are immediately descended from that node, separated by commas.
- Tips are represented by their names.
- Trees can be multifurcating at any level.
- Branch lengths can be incorporated into a tree by putting a real number after a node and preceded by a colon. This represents the length of the branch immediately below that node.

- Computer Representation of a phylogenetic tree

-Printing a phylogenetic tree

Printing a phylogenetic tree



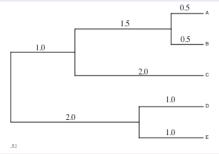
- Computer Representation of a phylogenetic tree

-Printing a phylogenetic tree

Printing a phylogenetic tree

Here are two examples:

rooted tree with edge lengths



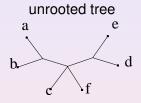
Newick string: ((((A:0.5,B:0.5):1.5,C:2.0):1.0,(D:1.0,E:1.0):2.0);

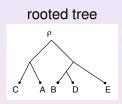
- Computer Representation of a phylogenetic tree

- Printing a phylogenetic tree

Printing a phylogenetic tree

Here are two more examples:





Newick string: ((a,b),c,f,(e,d)) or e.g.: ((a,b),(c,f,(e,d)))

Newick string: ((C,A),((B,D),E))

. . .

- Computer Representation of a phylogenetic tree

- Algorithm for printing a phylogenetic tree

Algorithm for printing a phylogenetic tree

The following algorithm recursively prints a tree in Newick format. It is initially called with e = null and $v = \rho$, if the tree is rooted, or v set to an arbitrary internal node, else.

- Computer Representation of a phylogenetic tree

-Algorithm for printing a phylogenetic tree

Algorithm for printing a phylogenetic tree

```
Algorithm toNewick(e, v)
     Input: Phylogenetic tree T = (V, E) on X, with labeling \lambda : V \to X
     Output: Newick description of tree
     begin
     if v is a leaf then
              print \lambda(v)
     else // v is an internal node
              print '('
              for each edge f \neq e adjacent to v do
                        If this is not the first pass of the loop, print ','
                        Let w \neq v be the other node adjacent to f
                        call to Newick(f, w)
             print ')'
     print ';
Stephanendele
```

- Computer Representation of a phylogenetic tree

-Parsing a phylogenetic tree

Parsing a phylogenetic tree

We need to be able to read a phylogenetic tree into a program. The following algorithm parses a tree in Newick format from a (space-free) string $s = s_1 s_2 \dots s_m$. The initial call is parseNewick(1, *m*, *null*).

- Computer Representation of a phylogenetic tree

-Parsing a phylogenetic tree

```
Algorithm parseNewick(i,j,v)
Input: a substring s_i \dots s_j and a root node v
Output: a phylogenetic tree T = (V, E, \lambda)
begin
while i \le j do
Add a new node w to V
if v \ne null then add a new edge \{v, w\} to E
```

if s_i = ' (' then // hit a subtree Set k to the position of the balancing close-bracket for i call parseNewick(i + 1, k - 1, w) // strip brackets and recurse

```
else // hit a leaf
Set k = \min\{k \ge i \mid s_{k+1} = `, ` or k + 1 = j\}
Set \lambda(w) = s_i \dots s_k // grab label for leaf
```

```
Set i = k + 1 // advance to next ', ' or j
if i < j then
Check that i + 1 \le j and s_{i+1} = ', '
Increment i // advance to next token end
```

- Computer Representation of a phylogenetic tree

-Parsing a phylogenetic tree

Parsing a phylogenetic tree

- In the Newick format, each pair of matching brackets corresponds to an internal node and each taxon label corresponds to an external node.
- To add edge lengths to the format, specify the length *len* of the edge along which given node we visited by adding :*len* behind the closing bracket, in the case of an internal node, and behind the label, in the case of a leaf.
- For example, consider the tree (a, b, (c, d)). If all edges have the same length 1, then this tree is written as (a:1,b:1, (c:1,d:1):1).

Drawing a phylogenetic tree

An **embedding** of a phylogenetic tree is given by an assignment of coordinates (x(v), y(v)) to each node v, such that

- ► any edge e = (v, w) is represented by the line segment connecting points (x(v), y(v)) and (x(w), y(w)),
- no two such line segments cross, and
- For every edge e, the length of the line segment representing e equals ω(e).

There are many different ways to draw a phylogenetic tree, both unrooted and rooted. We will describe a method for drawing an unrooted tree, called the **circular embedding** method.

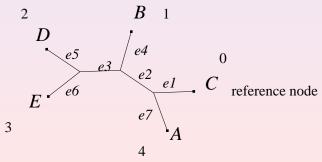
- Drawing a phylogenetic tree

- Computing a circular embedding

Computing a circular embedding

A circular embedding of a phylogenetic tree T is computed in two stages:

- 1. each edge *e* is assigned an angle $\alpha(e)$, and then,
- based on the edge angles and lengths, each node v is assigned coordinates (x(v), y(v)).



-Computing the edge angles

Computing the edge angles

Given a phylogenetic tree *T* with *n* leaves. Choose an arbitrary leaf *r* and call it the **reference** node. Each leaf *v* is assigned a **rank** $h(v) \in \{0, 1, ..., n-1\}$ defined as follows:

first set h(r) = 0 and then set h(v) = i, if v is the i-th leaf visited in a depth-first search (DFS)¹ of the tree from r.

We imagine the *n* leaves arranged around the unit circle in counter-clockwise order, with the *i*-th rank leaf *v* positioned at the angle $\alpha_i = \frac{i}{n} 2\pi$.

¹ Recall DFS: start from any node, go as far as possible (thus the name Depth First), until it can go no further. In this case, backtrack one step and repeat the process.

Stephan Steigele

Computing the edge angles

Computing the edge angles

Consider a fixed edge *e* and let *U* denote the set of all leaves that are separated from *r* by *e*. By definition of DFS, there exist two numbers p, q, with $1 \le p \le q \le n - 1$, such that $h(U) = \{p, p + 1, ..., q\}$. We define $\alpha(e)$ as the **mean angle** associated with the leaves that are separated from *r* by *e*:

$$\alpha(\boldsymbol{e}) = \frac{1}{\boldsymbol{q} - \boldsymbol{p} + 1} \sum_{i=p}^{q} \alpha_i = \frac{\pi}{n} (\boldsymbol{p} + \boldsymbol{q})$$

- Computing the edge angles

Computing the edge angles

This algorithm assigns an angle to every edge in the tree. The initial call is setAngles(0, null, r), where *r* is the reference leaf.

Algorithm setAngles(h, e, v)

Input: Number h of nodes placed, arrival edge e, current node v

Output: Angle $\alpha(e)$ for every edge $e \in E$.

Returns: New number of placed nodes.

begin

```
if v is a leaf and v \neq r then
```

return h+1

Set a = h // number of nodes placed before recursion

Set b = 0 // number of nodes placed after recursion

```
for all edges f \neq e adjacent to v do
```

```
Let w \neq v be the other node adjacent to f
Set b = \text{setAngles}(a, f, w)
Set \alpha(f) = \frac{\pi(a+b)}{n}
Set a = b
return b.
```

- Drawing a phylogenetic tree

- Computing the edge angles

Computing the edge angles

Lemma

The algorithm computes the edge angles of a circular embedding in linear time.

Computing the edge angles

Proof.

Every edge is visited exactly once, hence the runtime is linear in the number of edges |E| (which is linear in the number of leaves).

To see that the algorithm produces the correct result, consider the situation for some arbitrary edge e and node v.

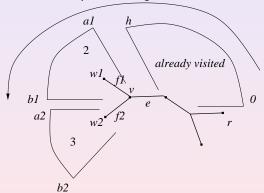
- The parameter h corresponds to the highest index assigned so far.
- ► Let f₁,..., f_k be the list of edges adjacent to v that are considered by the algorithm,
- let w_i denote the corresponding other node,
- and let a_i and b_i be the values of a and b directly after processing f_i.
- The latter two numbers denote the range of indices recursively assigned to the set of leaves in the subtree Stephan Steigel potted at w.

- Drawing a phylogenetic tree

Computing the edge angles

Computing the edge angles

The situation when processing node *v*:



We then compute

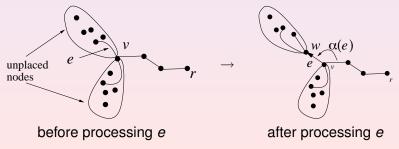
 $\alpha(f_i) = \frac{\pi(a_i+b_i)}{n} = \frac{2\pi}{n} \cdot \frac{a_i+b_i}{2} = \frac{2\pi}{n} \cdot \frac{a_i+a_{i+1}+\dots+b_i}{b_i-a_i+1}$, as required in the definition of a circular embedding.

- Drawing a phylogenetic tree

- Determining coordinates

Determining coordinates

Given a phylogenetic tree *T* and an angle function $\alpha : E \rightarrow [0, 2\pi]$, how do we obtain coordinates for the nodes? **Idea:** In a depth-first traversal of the tree starting at the reference node *r*, for every edge *e*, simply push the opposite node away from the current node in the direction specified by $\alpha(e)$ by the amount specified by the length $\omega(e)$:



- Drawing a phylogenetic tree

- Determining coordinates

Determining coordinates

The following algorithm does the pushing. Initially, we set $\epsilon(r) = (0,0)$ (the coordinates of the reference node *r*) and invoke setCoordinates(*null*, *r*).

```
Algorithm setCoordinates(e, v)

Input: Phylogenetic tree T and edge angles \alpha

Output: Circular embedding \epsilon of T

begin

Set p = \epsilon(v)

for each edge f \neq e adjacent to v do

Let w \neq v be the other node adjacent to f

Obtain p' by translating p in direction \alpha(f) by amount \omega(f)

Set \epsilon(w) = p'

call setCoordinates(f, w)

end
```

Drawing a phylogenetic tree

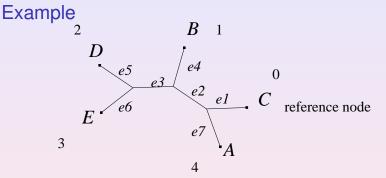
Determining coordinates

Determining coordinates

Theorem A circular embedding is computable in linear time. Challenge: prove that the resulting configuration is indeed a planar embedding, that is, that no two edges can cross.

- Drawing a phylogenetic tree

- Example



This is a circular layout with reference node C, which is verified as follows:

$$\alpha(e_1) = \frac{2\pi}{5\cdot 4}(1+2+3+4) = \pi$$

$$\alpha(e_2) = \frac{2\pi}{5\cdot 3}(1+2+3) = \frac{4}{5}\pi,$$

$$\alpha(e_3) = \frac{2\pi}{5\cdot 2}(2+3) = \pi,$$

Part II

Distance-Based Tree Reconstruction

-Outline

Distances Hamming Distance UPGMA **UPGMA** example The UPGMA Algorithm Application of the UPGMA algorithm Tree metrics The molecular clock hypothesis The ultrametric property Consistency Additivity and the four-point condition Neighbor-Joining The Neighbor-Joining algorithm Application of Neighbor-Joining Example Rooting unrooted trees

| Some | Topics | in Phy | logenetics |
|------|--------|--------|------------|
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-Outline

Constructing phylogenetic trees

There are three main approaches to constructing phylogenetic trees from molecular data.

- 1. Using a **distance method**, one first computes a distance matrix from a given set of biological data and then one computes a tree that represents these distances as closely as possible.
- 2. **Maximum parsimony** takes as input a set of aligned sequences and attempts to find a tree and a labeling of its internal nodes by auxiliary sequences such that the number of mutations along the tree is minimum.
- 3. Given a probabilistic model of evolution, **maximum likelihood** approaches aim at finding a phylogenetic tree that maximizes the likelihood of obtaining the given sequences.

Distances

Given a set $X = \{x_1, x_2, ..., x_n\}$ of taxa. The input to a distance method is a dissimilarity matrix $D : X \times X \to \mathbb{R}_{\geq 0}$ that associates a **distance** $d(x_i, x_j)$ with every pair of taxa $x_i, x_j \in X$. Sometimes we will abbreviate $d_{ij} := d(x_i, x_j)$ or $D_{ij} := d(x_i, x_j)$. We usually require that

- 1. the matrix is **symmetric**, that is, d(x, y) = d(y, x) for all $x, y \in X$, and
- 2. d(x, x) = 0 for all $x \in X$ and d(x, y) > 0 for all $x \neq y$, and
- 3. the triangle inequalities are satisfied:

$$d(x,z) \leq d(x,y) + d(y,z)$$
 for all $x, y, z \in X$.

-Hamming Distance

Hamming distance

Let a collection of taxa be given by a set of distinct sequences $A = \{a_1, a_2, \dots, a_n\}$ and assume we are given a multiple sequence alignment A^* of the sequences.

- We define sequence dissimilarity as the (normalized) Hamming distance Ham(a_i, a_j) between two taxa a_i and a_j as the number of mismatch positions in a^{*}_i and a^{*}_j, divided by the number of comparisons.
- We ignore any column in which both sequences contain a gap. If only one sequence has a gap in a column then we can either ignore the column, or treat it as a match, or as a mismatch, depending on the type of data.
- Usually, one ignores all columns in which any of the n sequences contains the gap.

-Hamming Distance

Hamming distance

Lemma

If the alignment is gap-less, then the corresponding distance matrix is a metric on A.

Proof.

Consider three distinct sequences $a_i, a_j, a_k \in A$. If $a_i^* \neq a_k^*$, then either $a_i^* \neq a_j^*$, or $a_k^* \neq a_j^*$, and hence $Ham(a_i, a_k) < Ham(a_i, a_j) + Ham(a_j, a_k)$.

-Hamming Distance

Hamming distance

For protein data, it makes sense to relax the definition of **sequence dissimilarity** to the number of "non-synonymous" residues divided by the number of sequence positions compared.

- For example, we may choose to ignore "conservative substitutions" by pooling amino acids with similar properties into six groups: acidic (D,E), aromatic (F,W,Y), basic (H,K,R), cysteine (C), non-polar (A,G,I,L,P,V), and polar (M,N,Q,S,T).
- Two residues are considered synonymous, if they are contained in the same group, and non-synonymous, otherwise.

- Distances

-Hamming Distance

Hamming distance

Example:

 a1
 C A A C C C C C A A A A A

 a2
 T A A T T T - C A A A A

 a3
 C G G T T T - - A A A A

 Distances:

$$Ham(a_1, a_2) = \frac{4}{12} = 0.\overline{33}$$
$$Ham(a_1, a_3) = \frac{5}{11} = 0.\overline{45}$$
$$Ham(a_2, a_3) = \frac{3}{11} = 0.\overline{27}$$

Hamming distances are only suitable for closely related sequences. Below we will discuss more sophisticated distances.

Question: What is the average Hamming distance between two Stephanrandom gap-free DNA sequences of the same length?

| Some | Topi | ics in | I Phyl | logen | etics |
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Distances

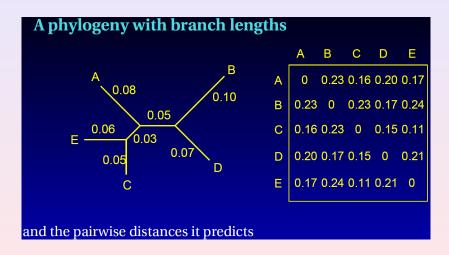
-Hamming Distance

Distance-Based Tree Reconstruction

Distance-Based Phylogeny Problem:

Reconstruct an evolutionary tree on *n* leaves from an $n \times n$ distance matrix **Input:** an $n \times n$ distance matrix $D = (d_{ij})$ and a labeling λ **Output:** a phylogenetic tree *T* (rooted or unrooted) with *n* leaves and edge lengths.

-Hamming Distance



Distances

-Hamming Distance

Least squares trees

Least squares methods minimize

$$\mathsf{Q} \; = \; \sum_{i=1}^n \sum_{j \neq i} \mathsf{w}_{ij} (\mathsf{D}_{ij} - \mathsf{d}_{ij})^2$$

over all trees, using the distances d_{ij} that they predict. Cavalli-Sforza and Edwards suggested $w_{ij} = 1$, Fitch and Margoliash suggested $w_{ij} = 1/D_{ij}^2$.

UPGMA

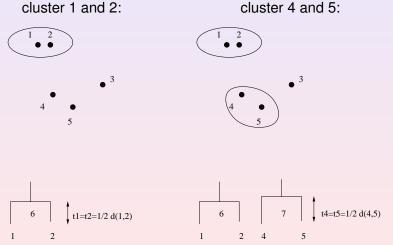
We will now discuss a simple distance method called UPGMA which stands for **unweighted pair group method using arithmetic averages** (Sokal & Michener 1958).

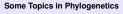
- Given a set of taxa X and a distance matrix D, UPGMA produces a rooted phylogenetic tree T with edge lengths.
- It operates by clustering the given taxa, at each stage merging two clusters and at the same time creating a new node in the tree.
- The tree is assembled **bottom-up**, first clustering pairs of leaves, then pairs of clustered leaves etc.
- Each node is given a height and the edge lengths are obtained as the difference of heights of its two end nodes.

-UPGMA

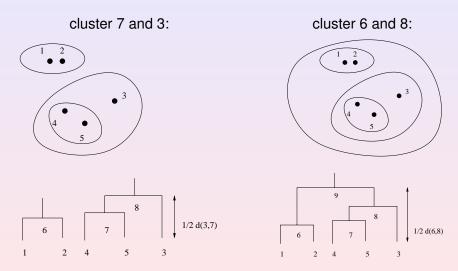
- UPGMA example

Example $X = \{1, 2, 3, 4, 5\}$, distances given by distance in the plane:





- UPGMA example



UPGMA produces a rooted, binary phylogenetic tree.

The UPGMA Algorithm

The distance between two clusters

Initially, we are given a distance d(x, y) between any two taxa, i.e. leaves, x and y. We define the **distance** $d(i, j) := d(C_i, C_j)$ between two clusters $C_i \subseteq X$ and $C_j \subseteq X$ to be the average distance between pairs of taxa from each cluster:

$$d(i,j) = \frac{1}{|C_i||C_j|} \sum_{x \in C_i, y \in C_j} d(x,y).$$

The distance between two clusters

Note that, if C_k is the union of two clusters C_i and C_j , and C_l is any other cluster, then

$$d(k, l) = rac{d(i, l)|C_i| + d(j, l)|C_j|}{|C_i| + |C_j|}.$$

This is a useful **update** formula, because using it in the algorithm, we can obtain the distance between two clusters in constant time.

The UPGMA Algorithm

The UPGMA algorithm

Algorithm UPGMA

Input: A set of taxa $X = \{x_1, ..., x_n\}$ and a corresponding distance matrix DOutput: A binary, rooted phylogenetic UPGMA tree $T = (V, E, \omega)$ on X

Initialization

Set $C = \{C_1 = \{x_1\}, \dots, C_n = \{x_n\}\}$ Assign each taxon x_i to its own cluster C_i Set $h(\{x_i\}) = 0$ //Define one leaf of T for each taxon, placed at height zero Set V = C and $E = \emptyset$

The UPGMA Algorithm

The UPGMA algorithm

Iteration

while $|C| \ge 2$ do Determine two clusters C_i and C_j for which d(i, j) is minimal Define a new cluster k by $C_k = C_i \cup C_j$ Set $C = (C - \{C_i, C_j\}) \cup \{C_k\}$ Set d(k, l) for all clusters l using the update formula Define a node k with daughter nodes i and j, and place it at height $h = \frac{d(i,j)}{2}$ Set $V = V \cup \{k\}$ and $E = E \cup \{(i, k), (j, k)\}$ Set $\omega(i, k) = h(k) - h(i)$ and $\omega(j, k) = h(k) - h(j)$

Termination

When only two clusters C_i and C_j remain, place the root at height $\frac{d(i,j)}{2}$.

- Application of the UPGMA algorithm

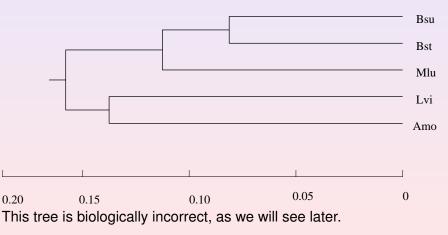
Example of UPGMA applied to 5S rRNA data:

| | | Bsu | Bst | Lvi | Amo | Mlu |
|---|-----------|-------|---------------|------------|----------|----------------|
| Original distances: | Bsu | _ | <u>0.1715</u> | 0.2147 | 0.3091 | 0.2326 |
| Abbreviations: | Bst | | _ | 0.2991 | 0.3399 | 0.2058 |
| Bsu: Bacillus subtilis Bst: Baclillus stearothermophilus | Lvi | | | _ | 0.2795 | 0.3943 |
| Lvi: Lactobacillus viridescens Amo: Acholeplasma modicum | Amo | | | | - | 0.4289 |
| Mlu: Micrococcus luteus | Mlu | | | | | - |
| | | B | su + Bst | Lvi | Amo | Mlu |
| | Bsu + B | st | _ | 0.2569 | 0.3245 | <u>0.2192</u> |
| \rightarrow | L | vi | | - | 0.2795 | 0.3943 |
| | An | 10 | | | - | 0.4289 |
| | М | lu | | | | — |
| | | | Bsu + E | Bst + Mlu | Lvi | Amo |
| Bs | u + Bst + | Mlu | | _ | 0.3027 | 0.3593 |
| , | | Lvi | | | _ | <u>0.2795</u> |
| | | Amo | | | | — |
| | | | В | su + Bst + | Mlu Lv | i + Amo |
| \rightarrow | Bsu + | Bst + | Mlu | — | <u>(</u> |). <u>3310</u> |
| | | Lvi + | Amo | | | - |

- Application of the UPGMA algorithm

Application of the UPGMA algorithm

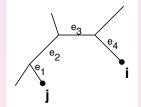
The resulting tree:



| Some Topics in Phylogenetics | |
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| | |
| Tree metrics | |

Tree metrics

Definition: Given a phylogenetic tree T on X. We say that a distance function $d : X \times X \to \mathbb{R}^{\geq 0}$ is **directly obtainable** from T, if each d_{ij} was obtained by adding up the edge lengths of the path between the leaves *i* and *j*.



 $d_{ij} = \omega(e_1) + \omega(e_2) + \omega(e_3) + \omega(e_4)$

Definition: Given a distance function $d : X \times X \to \mathbb{R}^{\geq 0}$ fulfilling the properties of a metric. Then *d* is a **tree metric** if there exists a phylogenetic tree *T* on *X*, such that *d* is directly obtainable from *T*.

| Some Topics in Phylogenetics |
|------------------------------|
| |
| Tree metrics |

Tree metrics

From this two fundamental questions arise:

- Is each distance function a tree metric?
- If d is a tree metric, does a unique tree and edge weight ω exist?

As we will see the answer to question one is "no", while positive answers for the second question exist.

| Some Topics in Phylogenetics | |
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| | |
| Tree metrics | |

Tree metrics

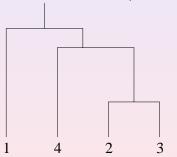
The goal of phylogenetic analysis is usually to **reconstruct** a phylogenetic tree from data, such as distances or sequences, that was produced by some **generating** or **true** tree.

- When reconstructing trees from real data, the generating tree is the path of events that evolution actually took.
- In this case, the true tree is unknown and the objective is, of course, to try and reconstruct it.
- In contrast, in simulation studies, a known tree T₀ is used to generate artificial sequences and/or distances, under some specified model of evolution.
- ► A tree reconstruction method is then applied and its performance can be evaluated by comparing the resulting tree *T* with the true tree *T*₀.

- The molecular clock hypothesis

The molecular clock hypothesis

Given a distance matrix *D*, the UPGMA method aims at building a rooted tree *T* with the property that all leaves have the same distance from the root ρ :



This approach is suitable for sequence data that has **evolved under** circumstances in which the rate of mutations of sequences is constant over time and equal for all lineages in the tree.

- UPGMA

The molecular clock hypothesis

The molecular clock hypothesis

Definition The assumption that evolutionary events happen at a constant rate is called the **molecular clock** hypothesis.

-UPGMA

The molecular clock hypothesis

UPGMA and the molecular clock

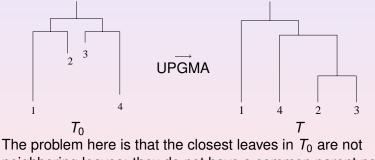
- If the input distance matrix D was directly obtained from a phylogenetic tree T₀ that fulfills the molecular clock assumption, then the tree T reconstructed by UPGMA from D will equal T₀.
- Otherwise, if T₀ does not do so, then UPGMA may fail to reconstruct the tree correctly

for example:

- UPGMA

- The molecular clock hypothesis

UPGMA and the molecular clock

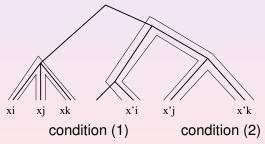


neighboring leaves: they do not have a common parent node.

- The ultrametric property

Definition: A distance matrix *D* is called an **ultrametric**, if for **every triplet** of taxa $x_i, x_j, x_k \in X$, the three distances $d(x_i, x_j)$, $d(x_i, x_k)$ and $d(x_j, x_k)$ have the property that either:

- 1. all three distances are equal, or
- 2. two are equal and the remaining one is smaller.



Lemma

if D was directly obtained from some tree T that satisfies the molecular clock hypothesis, then D is an ultrametric.

— The ultrametric property

The ultrametric property

We say that a rooted phylogenetic tree T is **ultrametric**, if every leaf has the same distance from the root. One can show the following result:

Theorem If D is a distance matrix directly obtainable from some ultrametric tree T, then UPGMA applied to D will produce that tree T. Consistency

Consistency

So, UPGMA computes the correct tree T, when given distances D = D(T) directly obtained from a ultrametric **model tree** T.

- In applications, we do not know T (of course) and thus cannot "directly obtain" D from T.
- ► We usually have an alignment of sequences and use it to compute an estimation D̂ of D(T).

Consistency

Given a suitable model of evolution (such as the Jukes-Cantor model discussed below) that specifies how the sequences evolve on T. As the length n of the aligned sequences increases, \hat{D} will converge to D.

A tree construction method is called **consistent** on a model tree *T*, if the tree that it produces "converges" toward *T*, as *n* increases.

Lemma

UPGMA is consistent on the set of ultrametic trees.

| Some Topics in Phylogenetics |
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| UPGMA |
| O |

Estimating the deviation from a molecular clock

Given a distance matrix D obtained by comparison of sequences generated along some unknown tree T_0 .

 Biologically, it may be of interest to know how well the molecular clock hypothesis holds.

In other words, how close is D to being an ultrametric?

Estimating the deviation from a molecular clock

To answer this question, we define the **stretch of an ultrametric** U with respect to D as follows:

$$stretch_{D}(U) = \max_{i,j \in X} \left\{ rac{D_{ij}}{U_{ij}}, rac{U_{ij}}{D_{ij}}
ight\}.$$

The **stretch of** *D* is defined as the minimum stretch over all possible ultrametrics:

stretch(D) = min_U{stretch_D(U)} and gives a lower bound for the stretch stretch_D(T) of any tree T obtained from D.

This value can be computed in $O(n^2)$ time, see ²

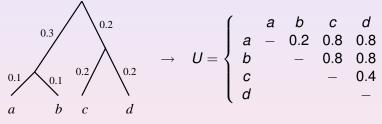
²L. Nakhleh, U. Roshan, L. Vawter and T. Warnow, LNCS 2452:287-299 (2002).

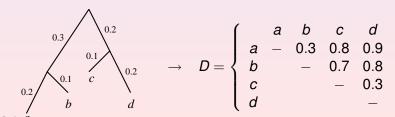
-UPGMA

Consistency

Estimating the deviation

Example of an ultrametric tree metric U and a non-ultrametric tree metric D:





| Some Topics in Phylogenetics |
|------------------------------|
| |
| Consistency |

Estimating the deviation from a molecular clock

The stretch of *U* w.r.t. *D* is:

$$stretch_D(U) = \max\left\{\frac{0.3}{0.2}, \frac{0.9}{0.8}, \frac{0.8}{0.7}, \frac{0.4}{0.3}\right\} = \frac{3}{2}.$$

- Additivity and the four-point condition

Additivity and the four-point condition

As we have seen a distance matrix often does not fulfill the ultrametric property. Now we ask if we can define a property of non-ultrametric tree metrics.

Definition: Given a set of taxa *X*. A distance matrix *D* on *X* fulfills the so-called **four-point condition** if for all $i, j, k, l \in X$

$$d(i,j) + d(k,l) \le \max\{d(i,k) + d(j,l), d(i,l) + d(j,k)\}$$

holds.

That is, two of the three expressions $d(x_i, x_j) + d(x_k, x_l)$,

 $d(x_i, x_k) + d(x_j, x_l)$, and $d(x_i, x_l) + d(x_j, x_k)$ are equal and are not smaller than the third.

- UPGMA

-Additivity and the four-point condition

Additivity and the four-point condition

The following theorem now tells us that the four-point condition and tree metric are equivalent properties, a result due to Peter Buneman (1971):

Theorem

A distance matrix D on X is a tree metric, if and only if it fulfills the four-point condition.

- UPGMA

- Additivity and the four-point condition

Additivity and the four-point condition

We want to at least prove one direction of the theorem:

- ▶ Let *D* be a tree metric on *X*.
- ► Then there exists a binary phylogenetic tree *T*.
- ► Let *i*, *j*, *k*, *l* be elements from *X*. If they are all different, then w.l.o.g we assume that (*i*, *j*) and (*k*, *l*) are neighbors respectively.

Then it follows that

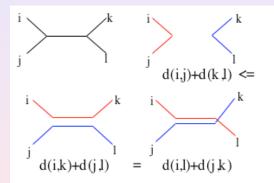
$$d(i,j) + d(k,l) \le d(i,k) + d(j,l) = d(i,l) + d(j,k)$$

as can be seen from the following figure:

- UPGMA

- Additivity and the four-point condition

Additivity and the four-point condition



and thus *D* fulfills the four-point condition. **Remark:** A tree metric is often called an **additive** metric.

- UPGMA

-Additivity and the four-point condition

The four-point condition

Example additive metric:

$$D = \left(\begin{array}{cccc} B & C & D \\ A & 7 & 6 & 5 \\ B & 3 & 6 \\ C & & 5 \end{array} \right)$$

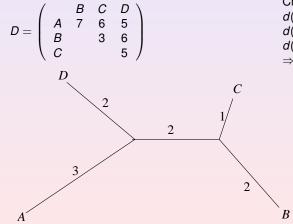
Check the four-point condition: d(A, B) + d(C, D) = 7 + 5 = 12 d(A, C) + d(B, D) = 6 + 6 = 12 d(A, D) + d(B, C) = 5 + 3 = 8 \Rightarrow the four-point condition holds.

- UPGMA

-Additivity and the four-point condition

The four-point condition

Example additive metric:



Check the four-point condition: d(A, B) + d(C, D) = 7 + 5 = 12 d(A, C) + d(B, D) = 6 + 6 = 12 d(A, D) + d(B, C) = 5 + 3 = 8 \Rightarrow the four-point condition holds.

- UPGMA

-Additivity and the four-point condition

The four-point condition

A non-additive metric:

$$D = \begin{pmatrix} B & C & D \\ A & 7 & 7 & 6 \\ B & 4 & 7 \\ C & 5 \end{pmatrix}$$

Check the four-point condition: d(A, B) + d(C, D) = 7 + 5 = 12 d(A, C) + d(B, D) = 7 + 7 = 14 d(A, D) + d(B, C) = 6 + 4 = 10 $\Rightarrow d(A, C) + d(B, D) \leq$ max (d(A, B) + d(C, D), d(A, D)) \Rightarrow 4-point condition doesn't hold.

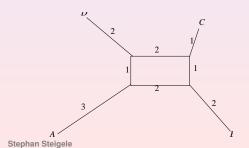
-UPGMA

Additivity and the four-point condition

The four-point condition A non-additive metric:

$$D = \begin{pmatrix} B & C & D \\ A & 7 & 7 & 6 \\ B & 4 & 7 \\ C & 5 \end{pmatrix}$$

Check the four-point condition: d(A, B) + d(C, D) = 7 + 5 = 12 d(A, C) + d(B, D) = 7 + 7 = 14 d(A, D) + d(B, C) = 6 + 4 = 10 $\Rightarrow d(A, C) + d(B, D) \leq$ max (d(A, B) + d(C, D), d(A, D)) \Rightarrow 4-point condition doesn't hold.



Neighbor-Joining

The most widely used distance method is **Neighbor-Joining (NJ)**, originally introduced by Saitou and Nei (1987)³, and modified by Studier and Keppler (1988).

- Given a distance matrix D, Neighbor-Joining produces an unrooted phylogenetic tree T with edge lengths.
- It is more widely applicable than UPGMA, as it does not assume a molecular clock.

³Saitou, N., Nei, Y. (1987) **SIAM J. on Comp.** 10:405-421 Stephan Steigele

Neighbor-Joining

Let D be a distance matrix directly obtainable from some (unknown) tree T.

Assume that we are building a tree based on *D* by repeatedly pairing "neighboring" taxa.

The following step reduces the number of leaves by one and we can repeatedly apply it until we arrive at a single pair of leaves:

- Let i and j be two neighboring leaves that have the same parent node, k.
- Remove *i*, *j* from the list of nodes and add *k* to the current list of nodes.

How do we have to set its distance to any given leaf m?

Neighbor-Joining

By **additivity of** *D*, we can compute the distances d_{km} from those between equivalent nodes in the original tree: k

In other words, for any three leaves i, j, m there is a node k where the paths to them meet.

Neighbor-Joining

By additivity,

$$d_{im} = d_{ik} + d_{km}, \ d_{jm} = d_{jk} + d_{km} \ \text{and} \ d_{ij} = d_{ik} + d_{jk},$$

thus

$$d_{im}+d_{jm}=d_{ik}+d_{km}+d_{jk}+d_{km}=d_{ij}+2d_{km},$$

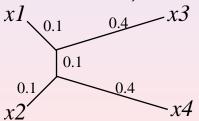
which implies $d_{km} = \frac{1}{2}(d_{im} + d_{jm} - d_{ij})$.

-Neighbor-Joining

Neighbor-Joining

How to determine which nodes are neighbors? The Neighbor-Joining method is based on the fact that we can decide which nodes are neighbors, using only the distance matrix.

However, it does not suffice simply to pick the two closest leaves, i.e. a pair *i*, *j* with *d_{ij}* minimal, for example:



Given distances generated by this tree.

Leaves x_1 and x_2 have minimal distance, but are not neighbors. Stephan Steigele

Neighbor-Joining

To avoid this problem, the trick is to **subtract the "averaged distances**"⁴ **to all other leaves**, thus compensating for long edges. We define:

$$N_{ij} := d_{ij} - (r_i + r_j),$$

where

$$r_i = \frac{1}{|L|-2}\sum_{k\in L}d_{ik},$$

and L denotes the set of leaves.

⁴Note that this is not precisely the average, as the number of summands is |L|, not |L - 2|.

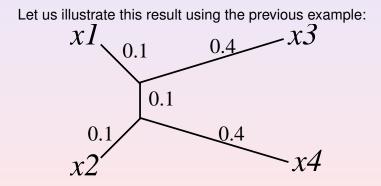
Neighbor-Joining

Lemma

If D is directly obtainable from some tree T, then the 2 leaves x_i and x_j for which N_{ij} is minimal are neighbors in T.

This result ensures that the Neighbor-Joining algorithm will correctly reconstruct a tree from its additive distances.

Neighbor-Joining



Neighbor-Joining

Here, $r_1 = 1/2(0.5 + 0.3 + 0.6) = 0.7$, and equivalently we compute $r_2 = 0.7$, $r_3 = 1.0$ and $r_4 = 1.0$. And so,

$$N = \begin{cases} x_2 & x_3 & x_4 \\ x_1 & -1.1 & -1.2 & -1.1 \\ x_2 & -1.1 & -1.2 \\ x_3 & & -1.1 \end{cases}$$

The matrix *N* attains a minimum value for the pair i = 1 and j = 3 and for the pair i = 2 and j = 4, as required.

-Neighbor-Joining

The Neighbor-Joining algorithm

Algorithm (Neighbor-Joining) Input: Distance matrix *D* Output: Phylogenetic tree *T* Initialization:

Define T to be the set of leaf nodes, one for each taxon.

Set L = T.

Iteration:

Pick a pair $i, j \in L$ for which N_{ij} is minimal. Define a new node k and set $d_{km} = \frac{1}{2}(d_{im} + d_{jm} - d_{ij})$, for all $m \in L$. Add k to T with edges of lengths $d_{ik} = \frac{1}{2}(d_{ii} + r_i - r_i)$ and

 $d_{ik} = d_{ij} - d_{ik}$, joining k to i and j, respectively.

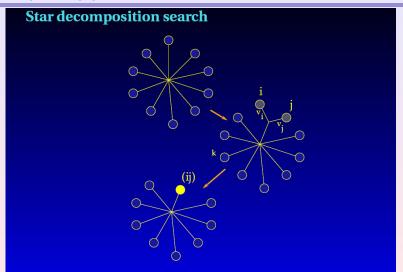
Remove i and j from L and add k.

Termination:

When *L* consists of two leaves *i* and *j*, add the remaining edge between *i* and *j*, with length d_{ij} .

-Neighbor-Joining

- The Neighbor-Joining algorithm



"Star decomposition" tree search method used in Neighbor-Joining method

The Neighbor-Joining algorithm

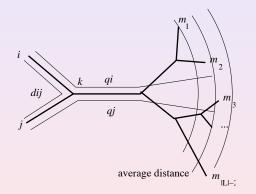
Neighbor-Joining algorithm

Why can we use $d_{ik} = \frac{1}{2}(d_{ij} + r_i - r_j)$ to update distances? By definition, $r_i = \frac{1}{|L|-2} \sum_{k \in L} d_{ik}$ equals the average distance $q_i = \frac{1}{|L|-2} \sum_{k \in L, k \neq i, j} d_{ik}$ from *i* to all other nodes $m \neq i, j$, plus $\frac{d_{ij}}{|L|-2}$:

-Neighbor-Joining

- The Neighbor-Joining algorithm

Neighbor-Joining algorithm



As we see from this figure:

$$2d_{ik} = d_{ij} + q_i - q_j = d_{ij} + q_i - q_j + \frac{d_{ij}}{|L|-2} - \frac{d_{ij}}{|L|-2} = d_{ij} + r_i - r_j.$$

-Neighbor-Joining

- Application of Neighbor-Joining

Application of Neighbor-Joining

- Given an additive distance matrix *D* directly obtained from a phylogenetic tree *T*, Neighbor-Joining is guaranteed to reconstruct *T* correctly.
- However, in practice we are never given a matrix that was "directly obtained" from the generating tree, but rather the distance matrix is usually obtained very indirectly by a comparison of finite sequence data generated along the tree. Such data is rarely additive.
- Nevertheless, the Neighboring-Joining method is often applied to such data and has proved to be a fast, useful and robust tree reconstruction method.

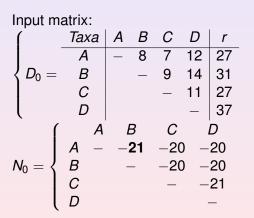
Moreover:

Lemma Neighbor-Joining is consistent.

-Neighbor-Joining

-Example

Example



 \rightarrow



- Example

Example

Data after one merge of neighbors:

$$D_{1} = \begin{cases} A+B & C & D & r \\ A+B & - & 4 & 9 & 13 \\ C & & - & 11 & 15 \\ D & & & - & 20 \\ N_{1} = \begin{cases} A+B & C & D \\ A+B & - & -10 & -7.5 \\ C & & - & -4.5 \\ D & & - & \end{cases}$$

 \rightarrow

| Some Topics in Phylogenetics |
|------------------------------|
| Neighbor-Joining |
| Example |
| |

Example

Data after two merges of neighbors:

$$D_{2} = \begin{cases} A + B + C & D & r \\ A + B + C & - & 8 \\ D & & - & 8 \\ 0 & & - & 8 \\ R \\ N_{2} = \begin{cases} A + B + C & D \\ A + B + C & - \\ D & & - \end{cases}$$

 \rightarrow

-Neighbor-Joining

-Example

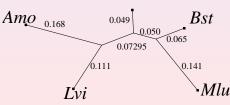
Example

Example of Neighbor-Joining applied to 5S rRNA data:

| | | Bsu | Bst | Lvi | Amo | Mlu |
|---|-----|-----|--------|--------|--------|--------|
| Original distances: | Bsu | _ | 0.1715 | 0.2147 | 0.3091 | 0.2326 |
| Abbreviations: | Bst | | _ | 0.2991 | 0.3399 | 0.2058 |
| Bsu: Bacillus subtilis Bst: Baclillus stearothermophilus | Lvi | | | _ | 0.2795 | 0.3943 |
| Lvi: Lactobacillus viridescens | Amo | | | | _ | 0.4289 |
| Amo: Acholeplasma modicum Mlu: Micrococcus luteus | Mlu | | | | | |

The resulting tree:

Bsu



- Rooting unrooted trees

Rooting unrooted trees

In contrast to UPGMA, most tree reconstruction methods produce an **unrooted** tree.

- Indeed, determining the root of a tree using computational methods is very difficult.
- In practice, the question of rooting a tree is addressed by adding an **outgroup** to the set of taxa under consideration.
- This is a taxon that is more distantly related to all other taxa than any other of the taxa.
- The root is then assumed to be on the branch attaching the outgroup taxon to the rest of the tree.

Rooting unrooted trees

Rooting unrooted trees

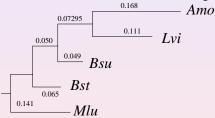
In practice, selecting an appropriate outgroup can be difficult:

- if it is too similar to the other taxa, then it might be more related to some than to others.
- If it is too distant, then there might not be enough similarity to the other taxa to perform meaningful comparisons.

Rooting unrooted trees

Rooting unrooted trees

In the above neighboring tree, Mlu is the outgroup, Hence, the rooted version of this tree looks something like this:

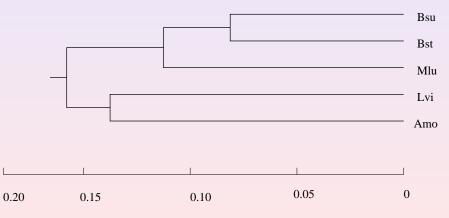


This tree is believed to be closer to the correct tree than the one produced earlier using UPGMA.

-Neighbor-Joining

- Rooting unrooted trees

Rooting unrooted trees



Rooting unrooted trees

Rooting unrooted trees

- In particular, the UPGMA tree does not separate the outgroup from all other taxa by the root node.
- The reason why UPGMA produces an incorrect tree is that two of the sequences, those of L.viridescens (Lvi) and A.modicum (Amo) are very much more diverged than the others.

Part III

Models of evolution

-Outline

Models of evolution

Jukes-Cantor Jukes-Cantor-Model Distance Transformations Accounting for superimposed events More general transformations

Models of evolution

In phylogenetic analysis, a **model of evolution** is given by a rooted tree T, called the **model**-, **true-** or **generating** tree, together with a procedure for generating sequences along the model tree.

- Usually, the procedure must determine how to generate an initial sequence at the root of the tree and specify how to "evolve" sequences along the edges of the tree.
- This involves obtaining intermediate sequences for all internal nodes of the tree, and producing a set of aligned sequences A at the leaves of the tree.

Observed and expected Distances

p-distance

Describes the proportion of different homologous sites, it is expressed as the **number of nucleotide differences per site**

g-distance

Accounts for the effects of homoplasy (e.g. $A \rightarrow G \rightarrow C$ or $A \rightarrow G \rightarrow A$) Here, the p-distance underestimates the true genetic distance.

Nucleotide Substitutions as homogeneous Markov Process

- Nucleotide Substitutions can be generalized as a Markov Process
- The relative rates of change of each nucleotide is specified in a Q-Matrix
- Rate of change is independent from one site to another (Markov Property)
- Substitution rates do not change (homogeneity)
- ► The relative frequencies of *A*, *C*, *G*, *T* are at equilibrium (stationary)

-Jukes-Cantor

The Jukes-Cantor model of evolution

T. Jukes and C. Cantor $(1969)^5$ *introduced a very simple model of DNA sequence evolution.* **Definition** Let *T* be a rooted phylogenetic tree. The **Jukes-Cantor** model of evolution makes the following assumptions:

- 1. The possible states for each site are A, C, G and T.
- 2. The sites evolve identically and independently (i.i.d.) down the edges of the tree from the root at a fixed rate *u*.
- With each edge e ∈ E we associate a duration t = t(e). The probabilities of change to each of the 3 remaining states are equal.

⁵T.H. Jukes and C.R. Cantor. (1969) Evolution of protein molecules. In Mammalian Protein Metabolism, H.N. Munro, ed., Academic Press, New York, NY, pp. 21-132.

-Jukes-Cantor-Model

Jukes-Cantor model

Under the Jukes-Cantor model, the evolutionary event that a nucleotide changes to any base occurs with a rate α .

- Let P and Q denote the base pair present at a given site before and after a given time period t.
- What is the probability Prob(Q = P | P, t) that P equals Q? (and hence become unchanged)

To answer this we make use of differential equations:



-Jukes-Cantor-Model

When to mutate ?

For t = 0 it is Prob(P|t = 0) = 1.

At the next time point t = 1 *P* either changes with rate α into one of the other 3 bases or it stays in the same state. Thus

$$Prob(Q = P | P, t = 1) = 1 - 3\alpha$$
.

Then it follows

$$Prob(Q = P|P, t = 2) =$$

$$(1 - 3\alpha)Prob(Q = P|P, t = 1) + \alpha(1 - Prob(Q = P|P, t = 1))$$

- Models of evolution

– Jukes-Cantor-Model

Exponential Function

After some transformations of the former equation we finally get the *exponential function* as solution. The probability that no mutation occured in the time interval (0, t).

 $P_0(t) = exp(-\mu t)$

The Probability of at least one event is then given by

$$P_1(t) = 1 - exp(-\mu t)$$

Note that the *Poisson Distribution* contains the exponential function.

$$P_{\lambda}(X=k)=rac{\lambda^k}{k!}\,\mathrm{e}^{-\lambda}$$

-Jukes-Cantor-Model

Probabilities of change from the **Q-Matrix**

A soon as the relative rates of change of each nucleotide are specified in a Q-Matrix it is possible to calculate the probability of change during evolutionary time t.

P(t) = exp(Qt)

Normally, detailed solutions require the use of matrix algebra, in case of Jukes-Cantor the result is straightforward:

-Jukes-Cantor-Model

Jukes-Cantor model

For Prob(P = P|P, t) we get the solution

$$\operatorname{Prob}_{ii}(t) = \frac{1}{4} + \frac{3}{4}e^{-\frac{4}{3}\alpha t}$$

while for $Prob(P \neq Q|P, t)$ we get the solution

$$\operatorname{Prob}_{ij}(t) = \frac{1}{4} - \frac{1}{4}e^{-\frac{4}{3}\alpha t}$$

For $t \to \infty \operatorname{Prob}(P|t)$ tends against 1/4. le., under the Jukes-Cantor model in equilibrium the probability of all four nucleotides is equal to 1/4.

-Jukes-Cantor-Model

Interpretation of the Jukes-Cantor model

Think about: how do we "evolve" a sequence down an edge *e* under the Jukes-Cantor model ?

- The evolutionary event (C₃ =) nucleotide change to one of the other three bases ocurs at a fixed rate u
- ► The event (C₄ =) nucleotide change to any base is taken to be ⁴/₃u = 4^u/₃
- ► The **probability** that event C_4 **does not** occur within time *t* is: $e^{-\frac{4}{3}ut}$ (Poisson distribution with k = 0)
- this leads also to a probablity of ¹/₄ to end in any particular base

- Models of evolution

-Jukes-Cantor-Model

Jukes-Cantor model

Let *P* and *Q* denote the base pair present at a given site before and after a given time period *t*. The probability that an observable substitution is detected (all changes from $P \in \{A, C, G, T\}$ to base *Q* within time *t* is:

$$\operatorname{Prob}(\boldsymbol{Q} \neq \boldsymbol{P} \mid \boldsymbol{P}, t) = \frac{1}{4} \left(1 - \boldsymbol{e}^{-\frac{4}{3}ut} \right).$$

However, there are three additional nucleotides, such that we have to sum over three such quantities to yield the **probability-of-change formula** for the probability of an **observable** change occurring at any given site in time *t*:

Prob(change
$$\mid t$$
) = $\frac{3}{4} \left(1 - e^{-\frac{4}{3}ut}\right)$.

-Jukes-Cantor-Model

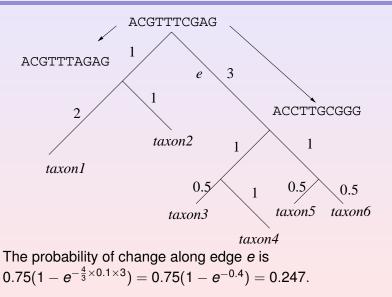
Jukes-Cantor model

We can use this model to generate artificial sequences along a model tree.

- E.g., set the sequence length to 10 and the mutation rate to u = 0.1.
- Initially, the root node is assigned a random sequence.
- Then the sequences are evolved down the tree, at each edge using the probability-of-change formula to decide whether a given base is to change:

-Models of evolution

-Jukes-Cantor-Model



- Models of evolution

Distance Transformations

The Jukes-Cantor distance transformation

- Given sequences generated under the Jukes-Cantor model of evolution.
- We would like to calculate the expected number of mutations for any given site on the path in a Jukes-Cantor tree between the leaves a_i and a_j.

Lemma

The maximum likelihood distance between a pair of sequences a_i, a_j (that is, the most likely ut to have generated the observed sequences) is given by the following formula:

$$JC(a_i, a_j) = -\frac{3}{4} \ln \left(1 - \frac{4}{3} Ham(a_i, a_j)\right).$$

This is called the **Jukes-Cantor distance transformation**.

-Accounting for superimposed events

Accounting for superimposed events

Hamming distances are suitable for inferring phylogenies between closely related species, given long sequences.

- For more distantly related sequences, the problem arises that mutation events will take place more than once at the same site.
- These superimposed events will not contribute to the Hamming distances and thus will go undetected.

-Accounting for superimposed events

Accounting for superimposed events

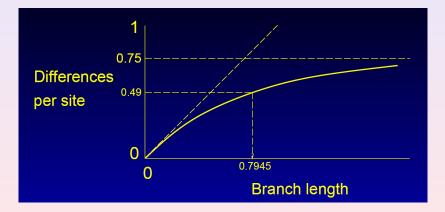
- ▶ Note that the expected Hamming distance between two random sequences is $\frac{3}{4}$. Any two sequences a_i, a_j for which $Ham(a_i, a_j) \ge \frac{3}{4}$ holds are called **saturated** w.r.t. each other.
- Note that the Jukes-Cantor transformation is undefined for any pair of saturated sequences.

In practice, when a saturated pair of taxa is encountered, their JC value is simply set to a large number which may be a fixed-factor times the largest value obtained between any two non-saturated taxa, for example.

- Models of evolution

-Accounting for superimposed events

genetic VS. observed distance



- More general transformations

More general transformations

The Jukes-Cantor model assumes that all nucleotides occur with the same frequency.

This assumption is relaxed under the Felsenstein 81 model introduced by Joe Felsenstein (1981), which has the following distance transformation:

$$F81(a_i, a_j) = -B\ln(1 - Ham(a_i, a_j)B),$$

where $B = 1 - (\pi_A^2 + \pi_C^2 + \pi_G^2 + \pi_T^2)$ and π_Q is the frequency of base Q.

The base frequency is obtained from the pair of sequences to be compared, or better, from the complete set of given sequences.

- More general transformations

More general transformations

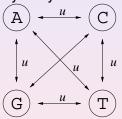
- Note that this contains the Jukes-Cantor transformation as a special case with equal base frequencies π_A = π_C = π_G = π_T = 0.25, thus B = ³/₄.
- ► Unlike the Jukes-Cantor transformation, this transformation can also be applied to protein sequences, setting $B = \frac{19}{20}$ if all amino acids are generated with the same frequency, and $B = \sum_{i=1}^{20} \pi_i^2$, in the proportional case.

- Models of evolution

- More general transformations

More general transformations

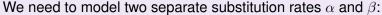
Both of these transformations assume that all changes of states are equally likely:

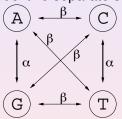


However, this is a very unrealistic assumption. For example, in DNA sequences, we observe many more **transitions**, which are purine-to-purine or pyrimidine-to-pyrimidine substitutions, than **transversions**, which change the type of the nucleotide.

- More general transformations

More general transformations





(Transitions: $A \leftrightarrow G$, $C \leftrightarrow T$, transversions: $A \leftrightarrow T$, $G \leftrightarrow T$, $A \leftrightarrow C$, and $C \leftrightarrow G$.)

More general transformations

More general transformations

Given equal base frequencies, but different proportions P and Q of transitions and transversions between a_i and a_j , the distance for the **Kimura 2 parameter** model is computed as:

$$K2P(a_i, a_j) = \frac{1}{2} \ln \left(\frac{1}{1 - 2P - Q} \right) + \frac{1}{4} \ln \left(\frac{1}{1 - 2Q} \right)$$

If we drop the assumption of equal base frequencies, then we obtain the **Felsenstein 84** transformation:

$$F84(a_i, a_j) = -2A \ln \left(1 - \frac{P}{2A} - \frac{(A - B)Q}{2AC}\right) + 2(A - B - C) \ln \left(1 - \frac{Q}{2C}\right),$$

where $A = \pi_C \pi_T / \pi_Y + \pi_A \pi_G / \pi_R$, $B = \pi_C \pi_T + \pi_A \pi_G$, and $C = \pi_B \pi_Y$ with $\pi_Y = \pi_C + \pi_T$, $\pi_R = \pi_A + \pi_G$.

- Models of evolution

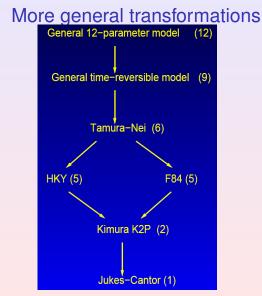
- More general transformations

More general transformations

Even more general models exist, but we will skip them. The following figure summarizes the complete overview of the time reversible models:

- Models of evolution

- More general transformations



Part IV

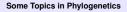
Trees and splits

-Outline

Definition of Split Systems

Trees and splits Compatible Splits Compatible splits and trees

Splits from a tree Splitstree

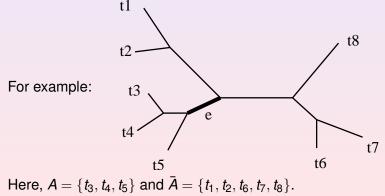


- Definition of Split Systems

- Trees and splits

Trees and splits

Any edge *e* of *T* defines a **split** $S = \{A, \overline{A}\}$ of *X*, that is, a partitioning of *X* into two non-empty sets *A* and \overline{A} , consisting of all taxa on the one side and other side of *e*, respectively.



- Definition of Split Systems

Trees and splits

Trees and splits

- We will use Σ(T) to denote the split encoding of T, i.e. the set of all splits obtained from T.
- If x ∈ X and S ∈ Σ, then we use S(x) or S(x) to denote the split part that contains x, or doesn't contain x, respectively.
- We define the size of a split S = {A, Ā} as size(S) = min(|A|, |Ā|).
- A split of size 1 is called a trivial split.

Ideally, each edge of the tree separates a monophyletic group from the rest and this is reflected by the corresponding split.

- Definition of Split Systems

- Compatible Splits

Compatible splits

Given a set of taxa X. Let Σ be a set of splits of X. Two splits $S_1 = \{A_1, \overline{A}_1\}$ and $S_2 = \{A_2, \overline{A}_2\}$ are called **compatible**, if one of the four following intersections

$$A_1 \cap A_2$$
, $A_1 \cap \overline{A}_2$, $\overline{A}_1 \cap A_2$, or $\overline{A}_1 \cap \overline{A}_2$,

is empty.

- Definition of Split Systems

- Compatible Splits

Compatible splits

A set Σ of splits of X is called **compatible**, if every pair of splits in Σ is compatible.

Example

- Given the taxa set $X = \{a, b, c, d, e\}$.
- ► The splits S₁ = {{a, b}, {c, d, e}}, S₂ = {{a, b, c}, {d, e}} and S₃ = {{e}, {a, b, c, d}} are all compatible with each other.
- ► However, S₄ = {{a, c}, {b, d, e}} is not compatible with the first one.
- Hence, the set $\Sigma = \{S_1, S_2, S_3\}$ is compatible, but $\Sigma' = \{S_1, S_2, S_3, S_4\}$ is not.

- Definition of Split Systems

Compatible Splits

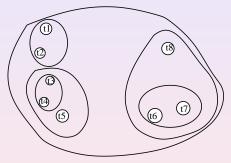
Compatible splits

- The compatibility condition states that any split S subdivides either the one side, or the other side, of any other split S', but not both sides.
- Hence, any set of compatible splits can be drawn as follows, without crossing lines:

- Definition of Split Systems

- Compatible Splits

Compatible splits



This figure also shows the relationship between compatible splits and a **hierarchical clustering**⁶

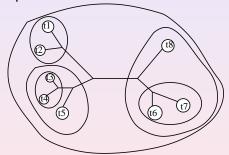
⁶A hierarchical clustering of a set *X* is a system \mathcal{H} of subsets of *X* such that $\bigcup_{A \in \mathcal{H}} A = X$ and $A, B \in \mathcal{H} \Rightarrow$ either $A \cap B = \emptyset$, or $A \subset B$ or $B \subset A$. Stephan Steigele

- Definition of Split Systems

- Compatible splits and trees

Compatible splits and trees

Any compatible set of splits Σ gives rise to a phylogenetic tree T, for example:



Note: To obtain a one-to-one correspondence between trees and compatible split systems, we now use the relaxed definition of a phylogenetic tree that allows any leaf to carry more than one label and also allows labeling of internal nodes, too.

- Definition of Split Systems

-Compatible splits and trees

Compatible splits and trees

In summary:

Theorem A set of splits Σ is compatible, iff there exists a phylogenetic tree *T* such that $\Sigma = \Sigma(T)$.

- Splits from a tree

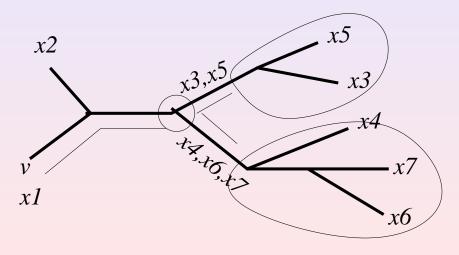
Splits from a tree

Given an unrooted phylogenetic tree *T* on $X = \{x_1, ..., x_n\}$. We can easily compute the set $\Sigma(T)$ in O(n), where *n* is the number of taxa,

- by starting at the leaf labeled x_1 ,
- visit all nodes in a post-order traversal,
- for each edge maintaining and reporting the set of leaves reached after crossing the edge.

-Splits from a tree

Splits from a tree



- Splits from a tree

Splits from a tree

The following algorithm recursively visits all nodes and prints out a split after visiting all nodes reachable over a particular edge. Initially, the algorithm is called with e = null and v equal to the node labeled x_1 .

```
Algorithm TreeToSplits(e, v)
Input: A phylogenetic tree T on X
Output: The corresponding compatible splits \Sigma(T)
Returns: The set \lambda(e) of all taxa separated from x_1 by e
begin Set \lambda(e) = \emptyset
for each edge f \neq e adjacent to v do
    Let w be the opposite node adjacent to f
    if w is a leaf then
      add the label of w to \lambda(e)
    else
      call TreeToSplits(f, w) and add the returned labels to \lambda(e)
print the split \{\lambda(e), \lambda(e)\}
return \lambda(e)
```

- Splits from a tree

A tree from splits

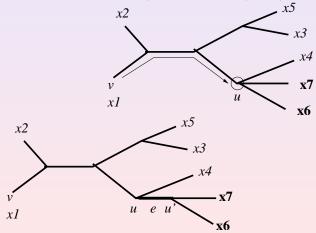
Given a compatible set of splits $\Sigma = \{S_1, S_2, \dots, S_m\}$ of *X*.

- Assume we have already processed the first *i* splits and have obtained a tree *T_i*.
- ► To incorporate a new edge *e* to represent the next split $S_{i+1} \in \Sigma$,
- starting at the node v labeled x₁, we follow a path through the tree
- until we reach the node w at which e is to be inserted

-Splits from a tree

A tree from splits

Example: insert split $\{x_1, x_2, x_3, x_4, x_5\}$ vs $\{x_6, x_7\}$:



- Splits from a tree

A tree from splits

This node is found as follows:

- if there is only one edge leaving the current node u that **separates** x_1 from elements in $\overline{S}(x_1)$, then we follow this edge.
- If more than one separating edges exist, then u = w
- create a new edge e from u to a new node u' and all separating edges are moved from u to u'.

- Splits from a tree

A tree from splits

Algorithm (Splits to tree) Input: A set $\Sigma = \{S_1, ..., S_m\}$ of compatible splits of X, (including all trivial ones) Output: The corresponding phylogenetic tree T = (V, E) on XInitialization: Let T be a star tree with n leaves labeled $x_1, ..., x_n$ Orient all edges away from the node v labeled x_1 For $e \in E$, maintain the set $\tau(e)$ of all taxa separated from x_1 by e

begin

for each non-trivial split $S \in \Sigma$ do

Set u = v

while there is only edge *e* leaving *u* with $\tau(e) \cap \overline{S}(x_1) \neq \emptyset$ do Set *u* equal to the opposite node of *e*

Let *F* be the set of all edges *f* leaving *u* with $\tau(f) \cap \overline{S}(x_1) \neq \emptyset$ Create a new edge *e* from *u* to a new node *u'*

Set $\tau(e) = \bigcup_{f \in F} \tau(f)$

Make all edges in F leave from w' instead of w

end

-Splits from a tree

A tree from splits

Lemma

The algorithm constructs the correct tree T in $O(n \log n)$ expected steps.

| Some | Topics | in Phy | logenetics |
|------|--------|--------|------------|
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-Splits from a tree

- Splitstree

Counting Splits

There are many possible splits

 $2^{n} - 2n - 2$

compared to the 2n - 3 splits that are compatible. For example: When n = 15 there are 27 *compatible* splits within 16,383 splits.

| Some Topics in Phylogenetics |
|------------------------------|
| Splits from a tree |
| Splitstree |
| |
| |

Splitstree

How to select for compatible splits?

- the Four-Point-Metric condition tells whether distances within a quartet of species fit within a tree
- applaying the FPM condition to all quartets of the given taxa does not guarantee that a unique tree implies all valid quartets

| Some Topics in Phylogenetics |
|------------------------------|
| Splits from a tree |
| Splitstree |

How to select for compatible splits?

A good try: Buneman index

- given the matrix D, Taxa X
- given split S = {A, B} of X, some x, y in A and some u, v in B

put

$$\beta(xy|uv) = \min\{d(x, u) + d(y, v), d(x, v) + d(y, u)\}$$
$$-(d(x, y) + d(u, v))$$

which defines the **Buneman index** $\beta_s = \frac{1}{2}min\beta(xy|uv)$

| Some | Topics | in Phy | logenetics |
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-Splits from a tree

Splitstree

Remarkable Facts: Buneman Index

Lemma

The collection of splits, for which $\beta_s > 0$ holds, is compatible and, therefore, corresponds to a tree.

However, the selection is too strict and usually elects to discard too many splits because only the minimum value is choosen and only one value has to be negative to reject the split.

| Some Topics in Phylogenetics | |
|------------------------------|--|
| - Splits from a troo | |

- Splitstree

Isolation Index

- given the matrix D, Taxa X
- given split S = {A, B} of X, some x, y in A and some u, v in B

put

$$\beta(xy|uv) = max\{d(x, u) + d(y, v), d(x, v) + d(y, u)\}$$
$$-(d(x, y) + d(u, v))$$

which defines the **Isolation index** $\alpha_s = \frac{1}{2} \min \alpha(xy | uv)$

| Some Topics in Phylogenetics | | |
|------------------------------|--|--|
| | | |

Consequences: Isolation index

- the Isolation Index corresponds to the weight of internal edges.
- however, the Splits S of the isolation index are not necesarily compatible
- ► Splits with positive isolation index are called *d* − *splits*

Lemma

if X has n elements, then the number of splits with positive isolation index is at most n(n-1)/2

| Some Topics in Phylogenetics |
|------------------------------|
| Splits from a tree |
| Splitstree |

Weak compatibility

all *d* – *splits* are **weakly compatible** Definition a split is *weakly compatible*, if for every *three* splits

$$S = \{A, B\}, T = \{C, D\}, U = \{E, F\}$$

at least one of the intersections

 $A \cap C \cap E, A \cap D \cap F, B \cap C \cap F, B \cap D \cap E$

is empty

| Some Topics in Phylogenetics |
|------------------------------|
| Splits from a tree |
| Splitstree |

Consequences: Weak compatibility

the FPM condition is also weakend by applying the weak compatibility

Lemma

when we look at the quartet ABCD we should have

 $max(D_{AB} + D_{DE}, D_{AE} + D_{DB}) > D_{AD} + D_{BE}$

for the quartet to be compatible with the split {ACDF|BEG}.

which means, that the distance sum of pairs of taxa within sets of the partition should not be the largets of the three sums

| Some Topics in Phylogenetics └─ Splits from a tree └─ Splitstree | |
|--|--|
| Drawing | |

The usual way to draw the networks is to utilize

outer planar networks.

However, details of the drawing algorithm are beyond the scope of this lecture.

Part V

Analysis and Benchmarking

-Outline

Comparison of two trees

Simulation studies

Algorithms vs optimality criteria

- Comparison of two trees

Comparison of two trees

Given two unrooted phylogenetic trees T_1 and T_2 on the same set of taxa *X*.

► We will call the first tree T₁ the model tree and T₂ the reconstructed tree.

How can we measure how similar their topologies are?

- Comparison of two trees

Comparison of two trees

- Let Σ₁ = Σ(T₁) and Σ₂ = Σ(T₂) be the split encodings of the two trees respectively.
- We define the set of all **false positives (FP)** as $\Sigma_2 \setminus \Sigma_1$.
- These are all splits contained in the estimated tree that are not present in the model tree.
- Similarly, we define the set of false negatives (FN) as
 Σ₁ \ Σ₂ as the set of all splits missing in the estimated tree, that are present in the model tree.

- Comparison of two trees

Comparison of two trees



Model TreeReconstructed TreeIn this example, there is one false positive $\{S_1, S_3\}$ vs $\{S_2, S_4, S_5\}$ and one false negative $\{S_1, S_2\}$ vs $\{S_3, S_4, S_5\}$.

-Simulation studies

Simulation studies

Simulation studies play an important role in bioinformatics. They are used to evaluate the performance of new algorithms on simulated datasets for which the correct answers are known.

- For example, to evaluate the performance of a tree construction method, *M*, we can proceed as follows:
 - 1. Select a model tree $T = (V, E, \omega)$ on X.
 - 2. For a specified sequence length *I* and mutation rate *u*, generate a set of aligned sequences *A* along the tree *T* under the Jukes-Cantor model.
 - 3. Apply the method M to A to obtain a tree M(A).
 - 4. Compute the number of false positives and false negatives.
 - 5. Repeat many times for many different parameters and report the average performance of the method.

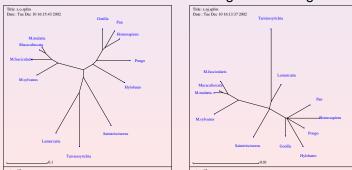
-Simulation studies

Simulation studies

Example Under the Jukes-Cantor model with L = 1000 and u = 0.1, sequences were generated on a model tree, then Hamming distances were computed, then **Neighbor-Joining** was applied:

Neighbor-Joining tree

FP - FN - 3

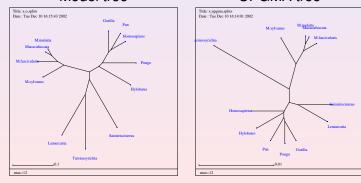


Model tree

Simulation studies

Simulation studies

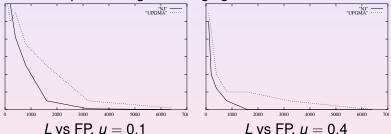
Example Under the Jukes-Cantor model with L = 1000 and u = 0.1, sequences were generated on a model tree, then Hamming distances were computed, then **UPGMA** was applied: Model tree UPGMA tree



FP = FN = 5

-Simulation studies

Using the same tree and computations, here we plot⁷ the number of false positives obtaining for u = 0.1, 0.5 and different sequence lengths *L* ranging from 100 - 6400:



For this particular tree, **Neighbor-Joining displays generally a better performance than UPGMA**. However, to obtain significant results, many trees and many different parameters <u>must be considered</u>.

⁷Each point plotted was obtained as the average of five independent simulations

-Simulation studies

Simulation studies

SeqGen: Program for simulation studies

Given the tree in Newick format: Generation of sequences using SeqGen⁸

⁸Rambaut, A. and Grassly, N. C. (1997) Seq-Gen: An application for the Monte Carlo simulation of DNA sequence evolution along phylogenetic trees. Comput. Appl. Biosci. 13: 235-238

- Algorithms vs optimality criteria

Algorithms vs optimality criteria

- In phylogenetics, the goal is to estimate the true evolutionary tree that generated a set of extant taxa.
- Tree reconstruction methods attempt to accomplish this goal by selecting one of the many different possible topologies.

- Algorithms vs optimality criteria

Algorithms vs optimality criteria

This selection process is performed in one of two ways:

- The tree selected by an **algorithmic** method such as Neighbor-Joining or UPGMA is defined by the sequence of steps that make up the algorithm. This tree does not solve any explicitly formulated optimization problem.
- 2. The tree selected by an **optimization** method is a solution to an explicitly formulated optimization problem such as "maximum parsimony" or "maximum likelihood". Here, algorithms play a secondary role as a means of obtaining or approximating a solution to the optimization problem.

- Algorithms vs optimality criteria

Algorithms vs optimality

Distance methods are usually **algorithmically** defined and have polynomial run time. Sequence methods usually involve solving an **optimization** problem by finding an optimal tree with respect to some criterion and are usually NP-hard.

- Algorithmic methods combine the computation and definition of the preferred tree into a single statement.
- Optimization methods involve formulating and solving two problems:
 - computing the cost for a given tree T, and
 - searching through all trees to find a tree that minimizes the cost.

Part VI Parsimony

| Some | Topi | cs in | Phyl | oge | enetics |
|------|------|-------|------|-----|---------|
|------|------|-------|------|-----|---------|

-Outline

Maximum parsimony

Assumptions for Maximum parsimony The parsimony score of a tree

Parsimony: Example

The small parsimony problem The Fitch algorithm Sankoff-Algorithm

The large parsimony problem

-Maximum parsimony

wi $\frac{1}{2}$ ga dem script

http://www-ab.informatik.unituebingen.de/teaching/ws02/abi1/AIBiI-WS2002-3-Huson.pdf

Maximum parsimony

Maximum parsimony

The maximum parsimony method is by far the most used sequence-based tree reconstruction method.

- In science, the principle of maximum parsimony is well known: always use the simplest, most parsimonious explanation of an observation, until new observations force one to adopt a more complex theory.
- In phylogenetic analysis, the maximum parsimony problem is to find a phylogenetic tree that explains a given set of aligned sequences using a minimum number of "evolutionary events".

-Maximum parsimony

-Assumptions for Maximum parsimony

Assumptions for Maximum parsimony

- Changes in different sites is independent.
- Changes in different lineages is independent.
- The probability of a base substitution that changes the amino acid sequence is small over the lengths of time in a branch.
- The expected amounts of change in different branches do not vary by so much that two changes in a high-rate branch are more probable than one change in a low-rate branch.
- The expected amounts of change do not vary enough among sites that two changes in one site are more probable than one change in another.

-Maximum parsimony

The parsimony score of a tree

The parsimony score of a tree

The **difference** between two sequences $x = (x_1, ..., x_L)$ and $y = (y_1, ..., y_L)$ is simply their non-normalized Hamming distance

$$\operatorname{diff}(x, y) = |\{k \mid x_k \neq y_k\}|.$$

-Maximum parsimony

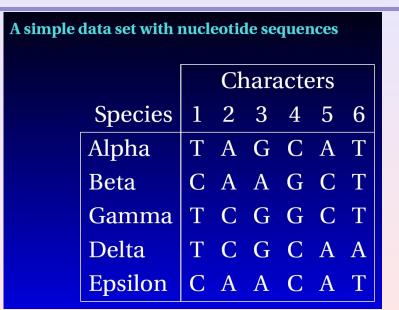
The parsimony score of a tree

The parsimony score of a tree

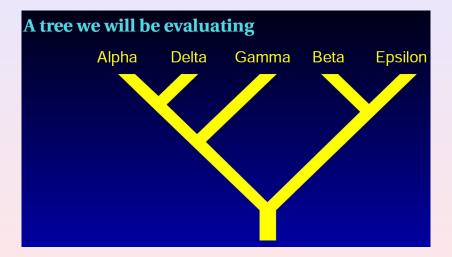
Given a multiple alignment of sequences $A = \{a_1, a_2, ..., a_n\}$ and a corresponding phylogenetic tree *T*, leaf-labeled by *A*.

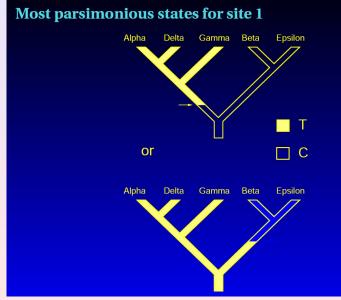
- If we assign a hypothetical ancestor sequence to every internal node in T,
- then we can obtain a score for T together with this assignment,
- by summing over all differences diff(x, y),
- where x and y are any two sequences labeling two nodes that are joined by an edge in T.

The minimum value obtainable in this way is called the **parsimony score** PS(T, A) of T and A.

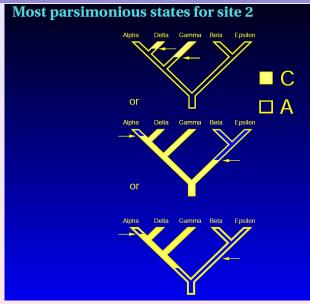


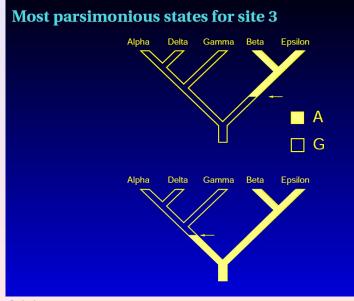
-Parsimony: Example

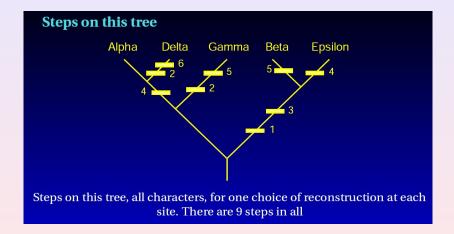


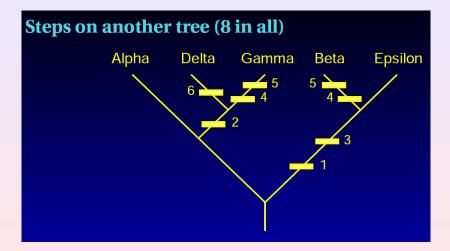


-Parsimony: Example







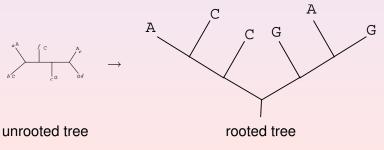


The small parsimony problem

The small parsimony problem

The **small** parsimony problem is to compute the parsimony score for a given tree T. **Can it be solved efficiently?**

As the parsimony score is obtained by summing over all columns, the columns are independent and so it suffices to discuss how to obtain an optimal assignment for one position:



- The small parsimony problem

The Fitch algorithm

The Fitch algorithm

- The following algorithm computes the parsimony score for T and a fixed column c in the sequence alignment.
- It uses a dynamic programming algorithm and assigns sets of character assignments S(v) to each node v in the tree.
- ► Initially it is called with e = null and v the root node and PS(T, c) = 0.

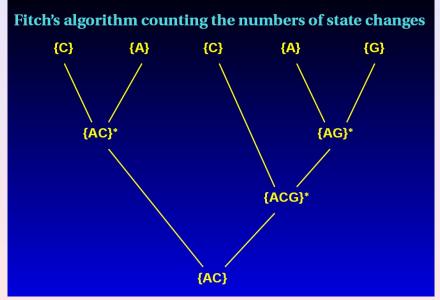
The small parsimony problem

The Fitch algorithm

The Fitch algorithm

```
Algorithm ParsimonyScore(e, v) (Walter Fitch 1971)
Input: A phylogenetic tree T and a character c(v) for each leaf v
Output: The parsimony score PS(T, c) for T and c
if v is a leaf node then
  Set S(v) = \{c(v)\}
else
  for each edge f_1, f_2 \neq e adjacent to v do
      Let w_i be the opposite node of f_i
      Call ParsimonyScore(f_i, w_i) // to compute S(w_i)
  if S(w_1) \cap S(w_2) \neq \emptyset then
    Set S(v) = S(w_1) \cap S(w_2)
  else
    Set S(v) = S(w_1) \cup S(w_2) and PS(T, c) = PS(T, c) + 1
end
```

- The small parsimony problem
 - The Fitch algorithm



- The small parsimony problem

The Fitch algorithm

Traceback

The above algorithm computes the parsimony score. An optimal labeling of the internal nodes is obtained via traceback:

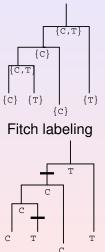
- starting at the root node r,
- we label *r* using any character in S(r).
- ► Then, for each child w, we use the same letter, if it is contained in S(w),
- otherwise we use any letter in S(w) as label for w.
- We then visit the children von *w* etc.

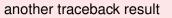
Again, the algorithm requires O(nL) steps, in total.

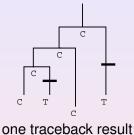
- The small parsimony problem

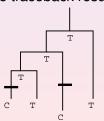
- The Fitch algorithm

Example:









not obtainable by traceback

- The small parsimony problem

The Fitch algorithm

The Fitch algorithm on unrooted trees

Above, we formulated the Fitch algorithm for rooted trees. However, the minimum parsimony cost is independent of where the root is located in the tree T. - The small parsimony problem

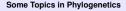
-Sankoff-Algorithm

more Advanced: Sankoff-Algorithm

A dynamic programming algorithm for counting the smallest number of possible (weighted) state changes needed on a given tree

- Let Sj(i) be the smallest (weighted) number of steps needed to evolve the subtree at or above node j, given that node j is in state i. Suppose that cij is the cost of going from state i to state j.
- Initially, at tip (say) j

$$S_j(i) = \begin{cases} 0 & \text{if node } j \text{ has (or could have) state } i \\ \infty & \text{if node } j \text{ has any other state} \end{cases}$$



- The small parsimony problem

-Sankoff-Algorithm

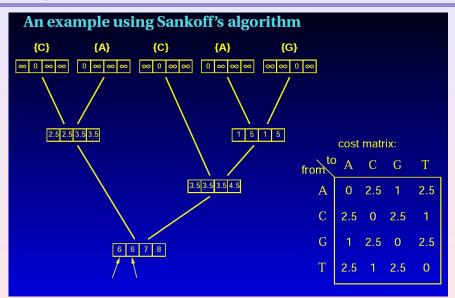
Then proceeding down the tree (postorder tree traversal) for node a whose immediate descendants are l and r

$$S_a(i) = \min_{j} [c_{ij} + S_l(j)] + \min_{k} [c_{ik} + S_r(k)]$$

The minimum number of (weighted) steps for the tree is found by computing at the bottom node (0) the S₀(i) and taking the smallest of these.

The small parsimony problem

-Sankoff-Algorithm



— The large parsimony problem

The large parsimony problem

Given a multiple alignment $A = \{a_1, ..., a_n\}$, its **parsimony** score is defined as

 $PS(A) = \min\{PS(T, A) \mid T \text{ is a phylogenetic tree on } A\}.$

The large parsimony problem is to compute PS(A).

- ▶ Potentially, we need to consider all (n-5)!! possible trees.
- Unfortunately, in general this can't be avoided and the maximum parsimony problem is known to be NP-hard.
- ► Exhaustive enumeration of all possible tree topologies will only work for n ≤ 10 or 11, say.

The large parsimony problem

The large parsimony problem

Thus, we need more efficient strategies that either solve the problem exactly, such as the branch and bound technique, or return good approximations, such as heuristic searches⁹.

⁹As with most biological problems, we are not only interested in the optimal solution, but would also like to know something about other near optimal solutions as well.

Part VII

Tree Searching Methods

-Outline

Branch and bound

Shortest Hamiltonian Path Problem But how to obtain an Branch and bound Algorithm for Phylogenetic Trees??

The stepwise-additional heuristic

The star-decomposition heuristic

Branch swapping methods

Nearest-Neighbor-Interchange (NMI) Subtree Pruning and Regrafting (SPR) Tree bisection and Reattachment (TBR)

Heuristic search

Simulated annealing

The Great Deluge method

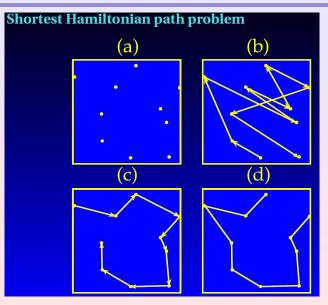
-Outline

Tree Searching Methods

- Exhaustive search (exact)
- Branch-and-bound search (exact)
- Heuristic search methods (approximate)
 - Stepwise addition
 - Branch swapping
 - Star decomposition

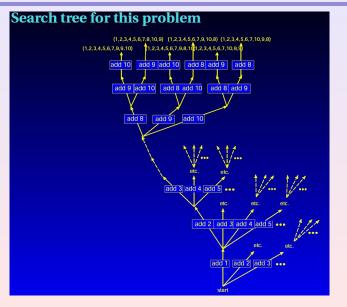
-Branch and bound

Shortest Hamiltonian Path Problem



-Branch and bound

-Shortest Hamiltonian Path Problem



-But how to obtain an Branch and bound Algorithm for Phylogenetic Trees??

But how to obtain an Branch and bound Algorithm for Phylogenetic Trees??

Recall how we obtained an expression for the number U(n) of unrooted phylogenetic tree topologies on *n* taxa:

- For n = 3 there are three ways of adding an extra edge with a new leaf to obtain an unrooted tree on 4 leaves.
- ► This new tree has (2n 3) = 5 edges and there are 5 ways to obtain a new tree with 5 leaves etc.

-But how to obtain an Branch and bound Algorithm for Phylogenetic Trees??

Branch and bound

- ► To be precise, one can obtain any tree T_{i+1} on {a₁,..., a_i, a_{i+1}} by adding an extra edge with a leaf labeled a_{i+1} to some (unique) tree T_i on {a₁,..., a_i}.
- In other words, we can produce the set of all possible trees on *n* taxa by adding one leaf at a time in all possible ways, thus systematically generating a complete enumeration tree.

-But how to obtain an Branch and bound Algorithm for Phylogenetic Trees??

Branch and bound

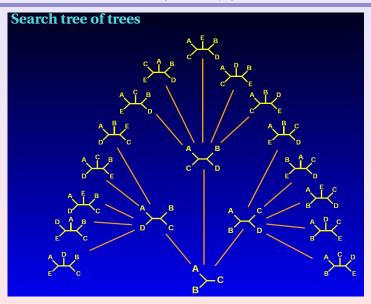
A simple, but crucial observation is that adding a new sequence a_{i+1} to a tree T_i to obtain a new tree T_{i+1} **cannot** lead to a smaller parsimony score.

This gives rise to the following bound criterion when generating the enumeration tree:

if the *local* parsimony score of the current incomplete tree *T'* is larger or equal to the best *global* score for any complete tree seen so far, then *we do not* generate or search the enumeration subtree below *T'*.

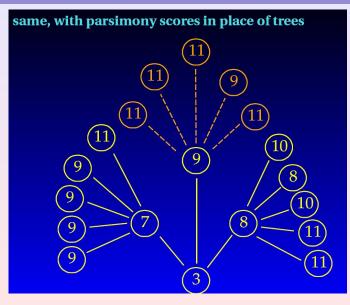
-Branch and bound

But how to obtain an Branch and bound Algorithm for Phylogenetic Trees??



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Branch and bound

- Application of branch-and-bound to evolutionary trees was first suggested by Mike Hendy and Dave Penny (1982).
- In practice, using branch and bound one can obtain exact solutions for data sets of twenty or more sequences, depending on the sequence length and the messiness of the data.
- A good starting strategy is to first compute a tree T₀ for the data, e.g. using Neighbor-Joining, and then to initialize the global bound to the parsimony score of T₀.

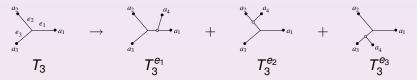
The stepwise-additional heuristic

The stepwise-additional heuristic

- Now we discuss a simple greedy heuristic (Felsenstein 1981) for approximating the optimal tree or score.
- We build the tree T by adding one leaf after the other, in each step choosing the optimal position for the new leaf-edge:
- Given a multiple sequence alignment $A = \{a_1, a_2, \dots, a_n\}$.
 - 1. Start with a tree T_2 consisting of two leaves labeled a_1 and a_2 .
 - Given T_i. For each edge e in T_i, obtain a new tree T^e_i as follows: Insert a new node v in e and join it via a new edge f to a new leaf w with label a_{i+1}.
 - 3. Set $T_{i+1} = \arg\min\{P(T_i^e, A)\}.$

The stepwise-additional heuristic

The stepwise-additional heuristic



- Obviously, this approach is not guaranteed to obtain an optimal result.
- Moreover, the result obtained will depend on the order in which the sequences are processed.

The star-decomposition heuristic

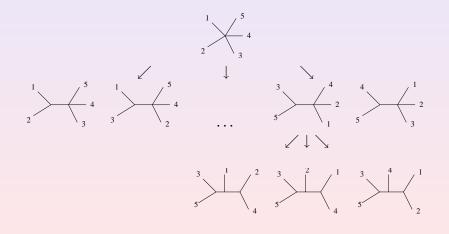
The star-decomposition heuristic

This employs a similar strategy to Neighbor-Joining.

- ▶ We start with a star tree on all *n* taxa.
- At each step, the optimality criterion is evaluated for every possible joining of a pair of lineages incident to the central node.
- The best tree found at one step is then used as the basis of the next step:

- The star-decomposition heuristic

The star-decomposition heuristic



- Branch swapping methods

Branch swapping methods

The two heuristics just described are both very susceptible to entrapment in local optima.

We now discuss a number of branch-swapping operations that one can use to move through the space of all trees, hopefully jumping far enough to escape from local optima.

- Branch swapping methods

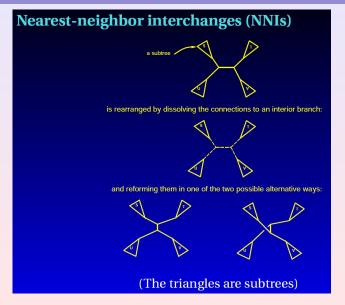
-Nearest-Neighbor-Interchange (NMI)

Nearest-Neighbor-Interchange (NMI)

In a **nearest-neighbor interchange (NNI)**, two of the four subtrees around an edge are swapped, in the two possible different ways:

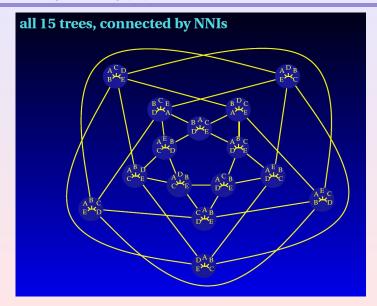
-Branch swapping methods

-Nearest-Neighbor-Interchange (NMI)



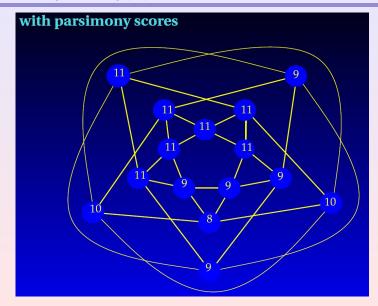
- Branch swapping methods

-Nearest-Neighbor-Interchange (NMI)



- Branch swapping methods

-Nearest-Neighbor-Interchange (NMI)



-Branch swapping methods

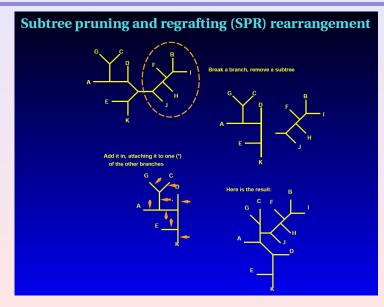
-Subtree Pruning and Regrafting (SPR)

Subtree Pruning and Regrafting (SPR)

In branch swapping by **subtree pruning and re-grafting**, a subtree is pruned from the tree and re-grafted to a different location of the tree:

-Branch swapping methods

-Subtree Pruning and Regrafting (SPR)

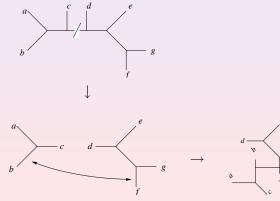


- Branch swapping methods

- Tree bisection and Reattachment (TBR)

Tree bisection and Reattachment (TBR)

In branch swapping by **tree bisection and reattachment** (TBR), the tree is bisected at an edge, yielding two subtrees. The two subtrees are then reconnected at two new positions:



-Branch swapping methods

Tree bisection and Reattachment (TBR)

Some Notes

- Each NNI operation is a special case of SPR operation, and each SPR operation is again a special case of TBR operation.
- We also note that each of the branch swapping operations is reversible such that if T' is the result of a branch-swapping operation on T then the same operation applied to T' yields back T.

-Branch swapping methods

Tree bisection and Reattachment (TBR)

Some Notes

In addition we have the following

Theorem

Let T and T' be two binary trees. Then T' can be constructed from T by a series of NNI operations. The number of different trees, that can be constructed from T with a single NNI operation is 2(n-3) and with SPR 4(n-3)(n-2). For the TBR the exact number of single operations depends on tree shape (but usually more than by SPR).

-Heuristic search

Heuristic search

If the data set $A = \{a_1, \ldots, a_n\}$ is too big to be solved exactly via branch and bound, then we can use a heuristic search method in an attempt to find or approximate the optimal solution.

- This involves searching through the space of all unrooted phylogenetic trees on *n* labels and trying to proceed toward a globally optimal one.
- We "move" through tree space using one or more branching-swapping techniques.

-Heuristic search

Heuristic search

Heuristic searches employ hill-climbing techniques:

- ▶ we imagine that the "goodness" PS(T) of the solution as a landscape along which we move during the search.
- The general strategy is to always move upwards in the hope of reaching the top of the highest peak.

Even using the above described branch-swapping techniques, any heuristic search is in danger of "climbing the wrong mountain" and getting stuck in a local optimum. Different strategies have been developed to avoid this problem. -Simulated annealing

Simulated annealing

The **simulated annealing** method employs a **temperature** that cools over time (Van Laarhoven and Aarts, 1987).

- At high temperatures the search can move more easily to trees whose score is less optimal than the score of the current tree.
- As the temperature decreases, the search becomes more and more directed toward better trees.

I.e., let T_i denote the current tree at step *i* and let $z(T_i)$ denote the goodness of T_i (e.g., -PS(T)).

-Simulated annealing

Simulated annealing

In hill climbing, a move to T_{i+1} is acceptable, if $z(T_{i+1}) \ge z(T_i)$. In simulated annealing, **any** new solution is accepted with a certain probability:

$$Prob(\text{accepting solution } T_{i+1})$$
$$= \begin{cases} 1 & \text{if } z(T_{i+1}) \ge z(T_i) \\ e^{-t_i(z(T_{i+1})-z(T_i))} & \text{otherwise,} \end{cases}$$

where *t_i* is called the **temperature** and decreases over time.

The Great Deluge method

The Great Deluge method

- The Great Deluge method, introduced by Guenter Dueck and Tobias Scheuer (1990), employs a slowly rising water level and the search accepts any move that stays above the water level.
- ► The probability of accepting a new solution *T_{i+1}* is 1, if *z*(*T_{i+1}*) > *w_i*, where *w_i* is a bound that increases slowly with time.
- If T_{i+1} is accepted, then we update the water level by setting

$$w_{i+1} = c \times (z(T_{i+1}) - z(T_i)).$$

► Typically, the constant *c* is usually about 0.01 to 0.05. Another of the many heuristics is **tabu search** method (Glover, 1989) that maintains a **tabu list** of 5 – 10 solutions recently visited and refrains from revisiting them.